Review

The Neurobiology of the Opsoclonus-Myoclonus Syndrome

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Summary: Opsoclonus-myoclonus is a pervasive neurological syndrome of children and adults. Although rare, it raises important clinical and neurobiological issues. This article provides an overview of the clinical and laboratory features, differential diagnosis, treatment, and outcome of opsoclonus-myoclonus. It pursues immunologic, genetic, electrophysiologic, neurochemical, and other clues to a pharmacologic model. Key questions include how and where the brain is injured, reversibility of the injury, possible targets for pharmacologic intervention, and which new studies are needed. Key Words: Myoclonus—Opsoclonus—Paraneoplastic—Neuroblastoma—ACTH.

The association of the ocular and somatic dyskinesias, opsoclonus and myoclonus, continues to tantalize pediatricians, oncologists, neuro-ophthalmologists, neurologists, movement disorder specialists, immunobiologists, molecular geneticists, and pharmacologists. Now more than 80 years since the original description of opsoclonus and its co-occurrence with myoclonus by Orzechowski (1,2), the syndrome of opsoclonus-myoclonus is internationally recognized as a neurologic manifestation of remote cancers and toxic, metabolic, infectious, structural, and degenerative disorders. Nearly 200 cases have been reported in children and adults. It has been variously named myoclonic encephalopathy (3-10) of infants (7,11-15) or childhood (16,17), dancing eyes (18,19), dancing feet (20), infantile polymyoclonia (21-23) or polymyoclonus syndrome (20,24-26), opsoclonus syndrome (27,28), acute cerebellar encephalopathy (29–32), encephalitis (33), or ataxia (34), syndrome of rapid irregular movements of eyes and limbs in childhood (35), oculocerebellomyoclonic syndrome (36-38), Kinsbourne syndrome (9,39-41), opsoclonus, body tremulousness, and benign encephalitis (42–43), syndrome of ocular oscillations and truncal myoclonus (44), encephalopathy associated with

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occult neuroblastoma (45), opsomyoclonus (46–48), or opsoclonus-myoclonus (49–55), opsoclonic cerebellopathy (56,57), or simply opsoclonus (58–60). The description opsoclonus, myoclonus, ataxia, (61) and encephalopathy (62) may be the most complete, but opsoclonus-myoclonus will be used here. There have been several large reviews of this syndrome (54,62–65), but none from the point of view of the movement disorder pharmacologist. A pharmacologic approach may be useful in identifying new hypotheses for study and potential new pharmacologic therapies.

CLINICAL FEATURES

Opsoclonus

Opsoclonus refers to conjugate or semiconjugate, chaotic, rapid, randomly directed eye movements, also called "saccadomania" (66). Although rare, opsoclonus may be dramatic. Orzechowski (1,2) said "the ocular globes are in a state of continuous agitation, being shaken and increasingly displaced by very rapid and unequal movements, which generally take place in the horizontal plane." Despite confusing terminology (67), the term opsoclonus is used by neuroophthalmologists in distinction from other ocular dyskinesias such as ocular myoclonus ("lightning eye movements") (68-70), ocular dysmetria (71), ocular flutter (71), and macrosaccadic oscillations (72). The relatedness of these movements is suggested by the occurrence of opsoclonus, ocular dysmetria, and ocular flutter in a pattern of temporal regression in the same patient (73). Orzechowski made the association of opsoclonus with ataxia and myoclonus. While opsoclonus is only one of several eye movement disturbances associated with myoclonus (74), myoclonus is the dyskinesia most often associated with opsoclonus. Opsoclonus may occur in "spells or bursts" (44,75-78). It persists with eyelids open or closed (77,79), but diminished (51). In sleep, opsoclonus may persist (11,12,76,80-82), though diminished (7,51), or may disappear (18-20). It is increased by saccadic movements (79) or fixation (12,44,51,72,76,78,81,83,84) and seldom decreased by fixation (43). Opsoclonus is increased by startle (12,81) or stimulation (78,85). Some patients prefer keeping one or both eyes closed (72,78,86), but for others, opsoclonus increases with eye closure (80). Oscillopsia has been reported (42,87, 88), but diplopia is absent (77,86).

Electronystagmyography (52,67) or electrooculography (35,87,89,90) has shown bursts of back-to-back saccades without saccadic interval in horizontal and vertical planes and dysconjugate features (78).

Opsoclonus may onset before the myoclonus (11,20,44,75). Opsoclonus may occur in the absence of myoclonus (91). In cases of coma, opsoclonus may persist (92). Occasionally, rotatory features have been noted (12,51,83,93) and what is described as opsoclonus is frequently called nystagmus (29,46,94). Opsoclonus may be increased by doll's-eye manuevers. Ice water calorics transiently interrupt (95), increase (81), superimpose deviation (11), or have no effect (44,77) on opsoclonus. Optokinetic nystagmus may be present (42,44,85) or absent (11,81).

Myoclonus

Although myoclonus has seldom been thoroughly described in any one report, it is possible to gain a collective impression from case reports (Table 1). The distribution of myoclonus may include the face (5,12,20,77,94,96,97), head and neck (5,12,27,75,78), limbs (18,20,27,35,51,62,77,78,85,98,99), fingers and hands (5,7,12,58,89), and the trunk (12,18,20,44,62,77,78) in truncal torsion jerks (11). Some authors comment that myoclonus was present in eyelids (12,18,100), or that there was eyelid fluttering or blinking (3,11,20,81,83,86,96). Some use the term "blepharospasm" (101,102). Palatal myoclonus is absent (53,77,100) except rarely (83). Respiratory impairment by myoclonus (12) or myoclonus of the diaphragm (83) is unusual. Unilateral myoclonus is also rare (94).

Myoclonus may occur spontaneously (12,51,103), but not always (53). It may be evoked by action (65,104) or intention (12,51,53). Other stimuli which induce myoclonus include noise, light, visual threat, and pinprick (11,12,35,53,77). Myoclonus is exacerbated by crying, excitement, or stress (12,18,20,27,65). Some cases are not stimulus-sensitive (86). The descriptions "asynchronous" or "irregular" have sometimes been applied (18,20,43,95,105–107). The term "minipolymyoclonus" (21,22,24,25,27,28) has been used to describe the small jerks that often involve only the fingers in opsoclonus-myoclonus, but implies primary generalized epileptic myoclonus (108). Not all jerks result in movement of a joint. There may be "attacks" of myoclonus (18). The severity of myoclonus is vari-

Feature	Apparent incidence ^a	Reference
Distribution		
Face	+ + + +	5, 12, 20, 77, 94, 96, 97
Eyelids	+ + +	12, 18, 100
Palate	-/+	83
Head and Neck	+ + +	5, 12, 27, 75, 78
Limbs	+ + + +	18, 20, 27, 35, 51, 62, 77, 78, 85, 98, 99
Fingers or Hands	+ + +	5, 7, 12, 58, 89
Trunk	+ + + +	12, 18, 20, 44, 62, 77, 78
Diaphragm	-/+	83
Туре		
Spontaneous	+	12, 51, 103
Action-induced	+ + +	65, 104
Intention-induced	+ +	12, 51, 53
Sensory-induced	+ + +	11, 12, 35, 53, 77
Light	+	
Sound	+ +	
Pinprick	+	
Emotion-exacerbated	+ + +	12, 18, 20, 27, 65
Functional Impairment		
Sitting	+ + + +	5, 11, 12, 18, 20, 29, 44, 51, 75, 77, 78, 83, 86, 98, 100, 101
Standing	+ + + +	5, 11, 20, 29, 42, 44, 51, 65, 77, 101, 104, 110, 111
Speech	+/-	5, 11, 12, 18, 29, 65, 71, 86, 110, 134
Feeding	+/-	11, 12, 137
Respirations	-/+	12

TABLE 1. Clinical description of myoclonus

a - / + indicates rare occurrence; + / - indicates infrequent occurrence.

able, ranging from violent (11,20,51,104) to occasional (80). There is no temporal association of myoclonus with opsoclonus (11,18,20,35,77,86). Myoclonus may onset before (18,77,103), or without opsoclonus (109). Myoclonus persists in sleep at slower rates (3,12).

The functional impact of myoclonus is typically severe. Standing and walking is usually compromised or refused (5,11,20,29,42,44,51,65,77,101,104,110,111). Involuntary kicking may occur when feet are placed on the ground (51). Patients are typically unable to sit (5,11,12,18,20,29,44,51,75,77,78,83,86,98,100,101) and revert to crawling (from walking) (11,12,40). Often, the patients "prefer lying down" (12,44,94), or "lying on back" (12), but may be unable to lie on back. One author comments the child cried when held upright (7).

Ataxia

The term "ataxia" has been used to mean apparently different things in the opsoclonus-myoclonus syndrome. Orzechowski (77) states that "among this entire picture of disease, we can always find a few symptoms of cerebellar disease . . ." Specific reference to "cerebellar ataxia" has been made (87,97,99, 108,112–115). Some patients were apparently ataxic enough to be discharged with a diagnosis of "acute cerebellar ataxia" (18,35,58,116), and "severe" ataxia has been described (5,7,44,51,62). There are reports of "titubation" of head or trunk (5,6,12,29,83), "truncal ataxia" (6,83,100,101,117,118), and limb dysmetria (29,65).

However, it has also been suggested that the "ataxia" is unlike cerebellar ataxia but is instead due to myoclonic jerks (20). Cerebellar function or finger-to-nose and heel-to-skin testing may be normal (43,44,75). Some reports of opsoclonus and ataxia make no mention of myoclonus (113). It is unclear if this is a biologic subgroup (119).

Early descriptions of the syndrome of "acute cerebellar ataxia of childhood" probably included some cases of opsoclonus-myoclonus. In a few, myoclonic movements (120,121), "trembling" (122), "jerking movements of the eyes" (122), or "action tremor" (123) were described. Even without those cases, however, there are more similarities than differences between the two syndromes, including a predilection for gait disturbance more than truncal or appendicular ataxia (120,121,123–125), encephalopathy, and behavioral and cognitive neurologic sequelae. There is enough overlap between opsoclonus-myoclonus and acute cerebellar ataxia syndromes so that children with the latter diagnosis should have urine catecholamine determinations made.

Tremor

Although tremor has been described in opsoclonus-myoclonus, the use of various other words or phrases has made it unclear whether true oscillatory tremor exists in this syndrome. "Intention tremor" has been described (6,11,18,43,58, 65,77,83,107), as well as "gross" or "coarse tremors" (27,42,51) and "shaking tremor" (86). Less clearly differentiating from fine myoclonic jerks are the terms "trembling" (11,117), "tremulousness" (43,52,80,97,101,126,127), "shakiness" or "quivering" (11,29), "shaking tremors" (77), and "shaking spells" (77). Associated intention tremors have also been equated to stimulus-sensitive myoclonus (78).

Encephalopathy

Mental and emotional features have been less well documented, but are suggested by use of such terms as anxiety (2), nervousness (102), lethargy (128), malaise (129), fretfulness (130), or irritability (20). Approximately half of the pediatric cases, regardless of etiology (65), are regarded as encephalopathic, but mental clouding is not usually a feature in children (12,77). Irritability may be dissociated from motor abnormalities which it may outlast (65).

In adults, encephalopathy ranges from irritability or mild emotional lability to coma and death (42,131). Altered mental status (apathy, lethargy, confusion) occurred in 58% of 19 adults with paraneoplastic opsoclonus (62,67,83,91,99,128, 123,133), and encephalopathy progressed to stupor or coma in 26% (91,99,128, 132). There was no apparent correlation of encephalopathy with other neurologic abnormalities, suggesting that it is not an obligate feature of the syndrome.

Other Neurologic Problems

Speech problems such as dysphasia, dysarthria with unintelligible speech, or mutism have been reported (5,11,12,18,29,65,71,110,134). Speech may also be normal (44,62,78).

Deep tendon reflexes are normal (20,43,44,78,100,104), decreased (11,53,86, 93), or increased (6,45,51,85,107). Babinski's sign may be present (58,62) or absent (43,44,107).

Muscle tone and strength are usually normal (45,51,104), but hypotonia (7,11, 12,20,45,53,100,106,135), which may be profound (3) and persistent (100), as well as increased tone (11,85) have been noted. A few patients are flaccid (58). Some patients have no head control (3). Only rarely has weakness been reported (36). Apparently, none of the patients have come to muscle biopsy.

Sensory examination is normal (44,45,53,77,85,86).

Other clinical features occur occasionally. Some older patients describe "dizziness" (44,78,100,110) or "vertigo" (62,128,136). Head nodding (20), urinary retention (53), drooling (11), or dysphagia (11,12,137) may occur. Focal or asymmetric features have been found. These include walking to one side (5), head tilt (12,138) or head deviation to one side (77), hemiparesis or unilaterally altered muscle tone (42,77), and asymmetric cerebellar signs (36). Hearing loss as an initial symptom occurs rarely (55). The diagnosis of posterior fossa neoplasm (12,26,29,124,139) or degenerative disease (12) is sometimes suggested. One patient lacked facial expression and tears (11).

DIFFERENTIAL DIAGNOSIS

Tumors outside the CNS and viral infections are the principal etiologies in children and adults, but other etiologies are not uncommon in adults. As many as

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half of the cases may be infectious in etiology (65). In pediatric cases, the mean age at onset is about 18 to 20 months (64,65). The youngest child with opsoclonusmyoclonus was apparently 4 months old (35,63). Only 13% of pediatric cases are older than 2 years (65). The age range of adult cases is broader, beginning usually with the third decade. Women are slightly more often affected than men (1.4:1) regardless of etiology. Non-neurologic prodromes occur in 36% within a month of onset of opsoclonus-myoclonus (64). These include upper respiratory or gastro-intestinal symptoms with equal frequency (63,140). A minority of cases received vaccinations within a month of neurologic signs (11,12,137–139). Neurologic symptoms may reach full expression earlier in nontumor-related cases, even within one week (65).

Infectious

Several different types of infections are associated with opsoclonus-myoclonus (Table 2) (2,11,12,42–44,51,80,85,96,101,102,141–156). A viral etiology may be the most common cause. The term "benign brainstem encephalitis" has been used (157,158). Viral prodromes of upper respiratory infections or gastroenteritis are typical, but do not rule out an underlying tumor (63). This is especially interesting in view of a proposed viral etiology of neuroblastoma (159). A small but definite group of viral pathogens has been identified from various body fluids of affected patients. Often, an infectious etiology can only be suspected (135,146,148).

Paraneoplastic Syndrome

Paraneoplastic opsoclonus-myoclonus (62,64,65,102,160) (Table 3) is distinct from other paraneoplastic disorders of the nervous system (161,162), such as cerebellar degeneration (163), myasthenic syndrome (Eaton-Lambert) (164), neuropathy and myopathy (165–167), encephalomyelitis (168), and limbic encephalitis (169). Paraneoplastic movement disorders are uncommon except for opsoclonus-

Agent	Reference	
Coxsackie $B_{3}^{A,B}$	51, 104	
Epstein-Barr ^{A,B}	80	
Hemophilus influenzae meningitis ^{A(B^a)}	85	
Herpes zoster ^{A,B}	141	
Immunization ^{A,B}	12, 154	
Lymphocytic choriomeningitis ^A	12	
Mumps ^A	44, 141, 143	
Neurosyphilis ^A	2	
Polioencephalitis ^A	141, 144, 145, 146, 147, 156	
$Psittacosis^{A(B^a)}$	101, 148	
Rubella ^A	11	
Salmonella typhi ^A	149	
St. Louis encephalitis ^{A,B}	43, 150, 151	
Tuberculous meningitis ^A	151, 96	
"Viral encephalitis" ^{A,B}	44, 47, 155, 146	

TABLE 2. Infectious etiologies of opsoclonus^A or myoclonus^B

^a Not all cases of opsoclonus were associated with myoclonus.

Clinical feature		Inf	ants			Ad	ults	
Reference	65		65		62		102	:
Number of cases	45		44		19		11	
Sex ratio (male:female)	0.6	5	0.7	,	1.	ł	0.	6
Mean age at onset (yr)	1.6	5 (0.5~5)			57.4	4 (29–77)	59.	1
Prodrome with vomiting	8	(18%)	10	(10%)	14	(74%)		
Onset of neurologic syndrome				. ,				
Within 1–2 wk	15 ^b	(56%)			12	(63%)		
After 1 wk	12 ^b	(44%)			6	(32%)		
Ataxia	45	(100%)	44	(100%)	18	(95%)	8	(72%)
Myoclonus	45	(100%)	35	(80%)	12	(63%)	5	(54%)
Opsoclonus	45	(100%)	41	(93%)	19	(100%)	11	(100%)
Dysarthria					4	(21%)		
Encephalopathy	19 ^a	(42%)			11	(58%)		
Course								
Complete recovery			15/2	5 (60%)				
Remitting-relapsing	10	(22%)	5/2	5 (20%)	4	(21%)		
2-year survival		(90%)			1	(<1%)		

TABLE 3. Opsoclonus-myoclonus as a paraneoplastic syndrome

^a Irritability.

^b Of 27 cases described in sufficient detail.

myoclonus. A syndrome of chorea changing to dystonia has been reported in an adult with small cell undifferentiated (oat cell) carcinoma and multiple medical problems (170).

There are a variety of associated neoplasms (Table 4) (3,4,7,20,26,27,29,36,46, 56,58–60,62,67,72,83,88,91,97–100,102,109,120,128,131,133,152,171–174). In children, neural crest-derived tumors predominate, such as neuroblastoma. "Gross nystagmoid movements of the eyeballs" were described in a child with neuroblastoma by Foster-Kennedy (175), but a direct connection between neuroblastoma and opsoclonus (58) and opsoclonus-myoclonus (176) was postulated later. In adults, the associated neoplasms are more heterogeneous. Some are also derived from neural crest cells, such as medullary thyroid carcinoma and oat cell carcinoma. However, many neural crest-derived tumors do not induce opsoclonus-myoclonus (Table 5). One such example is the pheochromocytoma, one of the more common pediatric endocrine neoplasms, which secretes pressor catechol-amines and induces many symptoms but not myoclonus.

Neuroblastoma originates from primitive sympathetic neuroblasts in the adrenal gland or sympathetic ganglion that do not differentiate (177). Neural crestderived tumors may be located throughout the body at any site along the pathway of cell migration (65,178,179). The neural crest-derived tumors are more often thoracic when associated with opsoclonus-myoclonus (49–61% mediastinal) (5, 65,180) then nonthoracic (46,181,182), but may originate at abdominalretroperitoneal (13%), adrenal (13%), sacrococcygeal (1%) or superior cervical ganglion locations (based on 23 cases). Neuroblastoma is the most common extracranial malignant neoplasm of early life (179). Ganglioneuroblastoma and ganglioneuroma occur one fifth and one tenth as often, respectively (183). In patients less than 14 years of age are found about 80% of neuroblastomas and 50% of ganglioneuromas (183). The ganglioneuroblastoma, with its different histologic

Primary tumor	Reference		
Bladder			
Anaplastic Ca ^{A,B}	132		
Brain			
Glioblastoma ^A	17, 171		
Pineal ^A			
Breast			
Infiltrating duct Ca ^{A,B}	62, 182		
Adeno Ca ^A	131		
Chondrosarcoma ^A	133		
Fallopian tube	133		
Adeno Ca ^A			
Lung			
Adeno Ca ^{A,B}	72		
Small cell ^{A,B}	62, 128		
Undifferentiated Ca ^{A,B}	56, 99		
Large cell ^{A,B}	67		
Epidermoid ^A	62		
Squamous cell ^A	172		
Lymphoreticular			
Lymphoma ^A	88		
Adrenal/sympathetic	20, 59		
Neuroblastoma ^{A,B}	3, 4, 26, 29, 58, 98		
Ganglioneuroblastoma ^{A,B}	7, 27, 29, 46, 58, 100, 173		
Ganglioneuroma ^{A,B}	109		
Sympathiconeuroblastoma ^A	36, 60		
Sympathicoblastoma ^A	120		
Pancreas			
Adeno Ca ^A			
Pharynx			
Squamous cell Ca ^A	56		
Thyroid			
Medullary Ca ^A	84		
Uterus			
Ca ^{A,B}	91, 174		

TABLE 4. Tumors associated with $opsoclonus^A$ or $myoclonus^B$

Ca = carcinoma.

subtypes (184) and sites of occurrence which affect prognosis (185), does occur in adults but apparently is not associated with opsoclonus-myoclonus.

Only 2-3% of neuroblastoma cases present as the paraneoplastic opsoclonusmyoclonus syndrome (28,177,182,186). A 0.5% incidence of occult neuroblastoma

Induces syndrome	Does not induce syndrome	
Neuroblastoma Ganglioneuroblastoma Ganglioneuroma Medullary thyroid carcinoma Oat cell carcinoma	Pheochromocytoma Islet cell tumor Carcinoid Paraganglioma Schwannoma Neurofibroma Merckel cell tumor Melanoma	

TABLE 5. Relation of neural crest-derived tumors to opsoclonus-myoclonus

has been reported in one autopsy series of infants less than 3 months of age (187). Spontaneous regression, which occurs by maturation into a ganglioneuroma or by cytolysis (177), is highest for neuroblastoma (188,189). Magnetic resonance imaging (190) and computed tomography (100,109,191–194) with or without ultrasound are more sensitive in detecting neuroblastoma than intravenous pyelogram (sensitivity 50%), plain radiography of chest or abdomen (40%), radionuclide bone scans (^{99m}Tc or ⁶⁷Ga) (50%), or physical examination (36%) (109,195–197). A 24-hour urine screen for catecholamines is routine. All of these measures may not detect the tumor. Bone marrow aspiration and skeletal surveys are not useful (65).

Failure to find a neuroblastoma does not preclude it as a cause of opsoclonusmyoclonus because it may be difficult to find (38,198) due to the possibility of spontaneous regression of the tumor (26,29,45,189). In 60% of cases, the tumor is found within 3 months (65). A delay in being able to diagnose neuroblastoma for up to four years after opsoclonus-myoclonus first appears may occur (7). The index of suspicion is so high that some patients without neuroblastoma have been followed for tumor for 12 years (35).

Rarely, opsoclonus-myoclonus may follow rather than precede removal of a neuroblastoma (6,199). Delayed onset of 15 months has been reported (199).

In adults, the instigating tumors are often local (56,62,67,72,102,128,173), but may be widely metastatic (62,133,200).

Reports of paraneoplastic opsoclonus without mention of myoclonus (56,58,62,87,91,96,102,133,149,171,172,201) are too frequent to dismiss. However, the absence of myoclonus or other dyskinesias that might actually have been myoclonus is seldom documented (100).

Unlike adults, children with tumor-associated opsoclonus-myoclonus have an excellent prognosis regarding survival: 90% two-year survival rate compared to 30-34% in other patients with neuroblastoma (46,65,182,202–205). Earlier diagnosis and lower tumor stages (206,207) at the time of diagnosis only partially account for improved prognosis. However, some of the tumors are metastatic (7,29,36) and deaths have been reported (7,29,36,120,208). Some of these deaths apparently have been postsurgical complications (7) or, rarely, due to inoperability (120).

Opsoclonus-myoclonus also is frequently associated with pervasive and permanent neurologic and cognitive deficits even after the tumor is removed surgically (16). Psychomotor retardation may persist even when opsoclonusmyoclonus abates (26,29,31,58).

Relapses of opsoclonus-myoclonus are associated with intercurrent illnesses, tumor therapies, changes in medications, and other factors (65). The clinical course may chronically fluctuate (12,18,24,65,116,209) or spontaneously remit (65,96,130,155).

Neuroblastoma may be associated with other neurologic syndromes besides opsoclonus-myoclonus involving the peripheral (Horner's syndrome and palsies of peroneal, phrenic, recurrent laryngeal, or facial nerves or Erb's palsy) or central nervous system (intracerebral, intraspinal) (172,182,210). Occasionally, paraneoplastic opsoclonus-myoclonus may occur with other central involvement such as intradural extension of an abdominal retroperitoneal neuroblastoma (199).

Toxic-Metabolic

Many drugs induce myoclonus at toxic or pharmacologic doses, fewer drugs evoke opsoclonus, and the co-occurrence of drug-induced opsoclonus and myoclonus is uncommon (87,105,153,210,211–260) (Table 6). Some cases are reversible, such as the stimulus-sensitive action or intention myoclonus induced by tricyclic antidepressants (261), whereas others are not. Ketamine hydrochloride reversibly exacerbated opsoclonus and myoclonus in an infant with neuroblastoma-associated opsoclonus-myoclonus (40).

While myoclonus is a feature of several different inborn errors of metabolism, few are associated with opsoclonus. The syndrome of opsoclonus-myoclonus has been observed in an adult with hyperosmolar nonketotic coma (52,95) and a child with multiple carboxylase deficiency (262).

Other Conditions

Opsoclonus occurs in apparently normal newborns (76,263) and rarely (with or without myoclonus) in association with congenital malformations, vascular genetic, metabolic, degenerative, and other acquired disorders (2,52,71,81, 95,262,264–281) (Table 7). Unusual associations of opsoclonus-like movements have also been described (66,91,282).

LABORATORY TESTS

Electroencephalography

Electroencephalography is commonly performed in cases of opsoclonusmyoclonus. Most encephalograms (EEGs) have been normal (3,5,11,12,18,20, 27,29,35,40,43,45,51,89,100,103,190). No epileptiform activity has been reported except in meningitis (85). Slowing, almost always diffuse, usually mild but occasionally severe is sometimes seen in both infants (45,58) and adults (19,62,67,68,78,86,91,95,99,102,218). The EEG may normalize during steroid treatment, but no causal relation has been established.

Clinical seizure activity is extremely rare. In the few reported cases, only a few seizures occurred (45,283,284).

Evoked Potentials

Normal (45,285) as well as abnormal (286) brainstem auditory, visual-evoked and somatosensory responses have been reported. Three children with opsoclonus-myoclonus had mildly abnormal brainstem auditory evoked potentials which were interpreted as indicating tegmental pontine lesions affecting the lateral lemniscus and brachium conjunctivum (106). Delayed wave V and prolonged interpeak latencies were found.

Electromyography

Electromyography (EMG) may be used to confirm the myoclonic nature of the dyskinesias (15). EMG bursts are typically brief (20-60 msec) (35). EMG has

shown independent, asynchronous myoclonic jerks at rest, aggravated by voluntary movements (89), with a silent period (19). No correlation has been found between electroencephalography and EMG activity on routine testing (27).

Neuroimaging

Two adults with opsoclonus-myoclonus following an upper respiratory illness were found to have pontine tegmental lesions on magnetic resonance imaging (MRI) (53). One of the cases had visual hallucinations, a lesion at the junction of the pontine basis and tegmentum, and an old lacunar infarction in the right putamen (also shown on cranial computed tomography, CT). However, normal head MRI studies have also been reported in adults (62,102,285). A questionable T_2 hyperintensity in the central and left posterior midbrain was reported in an adult with opsoclonus (287).

Cranial CT scans in most cases of opsoclonus-myoclonus have been normal in children. A low-density cerebellar lesion posterior to the fourth ventricle has been reported (97). One child with opsoclonus-myoclonus had a lesion in the cerebellar vermis (288).

Early studies reported normal skull series (42,43,77,100,101,174), brain scans (100,101,172), and cerebral angiography (56,91,275). Cerebellar atrophy has been noted infrequently on pneumography (27,29,116), but most pneumoencephalograms have been normal (19,71,172).

Cerebrospinal Fluid Studies

Lumbar punctures are usually performed in the acute phase of the illness. Cerebrospinal fluid (CSF) may be normal (11,12,18,27,29,40,45,56,77,78,89, 103,107,113), show isolated pleocytosis (3,18,19,26,42,44,45,51,90,120,149), or pleocytosis with increased protein in intracranial tumor (171) or encephalitis (43). CSF glucose is typically normal. In adults with a paraneoplastic etiology, CSF shows lymphocytic pleocytosis often with slight protein elevation (53%) (62).

CSF immunoglobulins may be normal (18,27), but increased IgG and IgM have been found in a few cases of opsoclonus in CSF (20,80,85,106,172) and serum (172).

CSF oligoclonal bands have been reported (62,80,102,104,106,107).

In cases of extracranial neuroblastoma without opsoclonus-myoclonus, increased lumbar CSF homovanillic acid (HVA) compared to other extracranial tumor controls has been reported (289). HVA, hydroxymethoxyphenyl ethylene glycol (HMPG) and vanillylmandelic acid (VMA) were elevated in six patients with intracranial or cranial neuroblastoma (mean age 5 years) compared to 16 patients (mean age 2 years) with extracranial neuroblastoma. However, controls were not age-matched (mean age 17–33 years), and it is well known that lumbar CSF catecholamine levels are higher in infants than children, which are, in turn, higher than adults.

There has only been one report of CSF neurotransmitter metabolites in opsoclonus-myoclonus (290). In a study of 10 patients, CSF 5-hydroxyindoleacetic acid (5-HIAA) and HVA were decreased in patients with opsoclonus-myoclonus

Substance	Reference
Amitriptyline ^{A,B}	212, 213, 214
Chlordecone ^{A,B}	215
Cocaine (intranasal) ^{A,B}	216
DDT ^{A,B}	217
Diazepam ^A	87
Lithium-haldol ^A /Lithium ^B	218, 219, 220
Organophosphates ^{A,B}	105, 221
Phenytoin ^{A,B}	87
Thallium ^A	201
Toluene ^A	222
Alcohols ^B	229
Amphetamine ^B	229
Apresoline ^B	229
Apresonne Bemegride ^B	105
Bemegride ^B Benzamides ^B	252
Biguanides ^B	232
Bismuth salts ^{$B(A^a)$}	
Borates ^B	223, 224, 225 229
Borales ⁻	229
Buflomedil ^B	
Carbamazepine ^B	227, 228, 251
Camphor ^B	105
Cefmetazole ^B	257
Chloralose ^B	235
Chlorambucil ^B	230
Clomipramine ^B	253
Codeine ^B	229
Cycloserine ^B	105
Dextropropoxyphene ^B	229
Dibenzoxazepines ^B	258
Enflurane anesthesia ^B	231
Ethylene glycol ^B	229
Etomidate anesthesia ^B	232, 249, 250
Fentanyl ^B	245
Fluoracetate ^B	105
Hydrazides ^B	232
Imipramine ^B	234
Insulin ^B	229
Isoniazid ^B	153, 229
L-dopa (parkinson patients) ^B	235, 236
Lead ^B	237
Menthol ^B	229
Mercury and its salts ^B	237
Metaldehyde ^B	229
Methaqualone ^B	238
Methyl bromide ^B	239
Metoclopramide ^B	240, 259
Morphine ^B	241
Naloxone ^B	256
Nicotine ^B	229
Nikethamide ^B	229
Norpethidine ^B	242
Paradichlorobenzene ^B	229
Penicillin ^B	242
Pentetrazole ^B	244
Phencyclidine ^B	246

TABLE 6. Drugs or toxins associated with $opsoclonus^A$ or $myoclonus^B$

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(continued)

Substance	Reference
Phenelzine ^B	252
Phenol ^B	105
Picrotoxin ^B	229
Piperazine ^B	247
Propfenone ^B	260
Sodium santonin ^B	237
Strychnine ^B	281
Sufentanil ^B	255
Sulfamides ^B	229
Theophylline ^B	229
Tris(hydroxymethyl)aminomethane ^B	229

TABLE 6. Continued

^a Described as atypical opsoclonus.

who were 4 years old or younger but not in older children compared to 21 age- and sex-matched controls. None of the patients exhibited increased CSF 5-HIAA. No clinical differences were seen in the subgroup with low CSF 5-HIAA.

The presence of interferon was reported in the CSF of a child with opsoclonusmyoclonus of presumed viral etiology (291).

Tumor Markers and Growth Factors

Both cellular and circulating markers for neuroblastoma have been described (292). Cellular markers include antigens detected by monoclonal antibodies, oncogenes, and radiolabelled MIBG ([131 I]meta-iodobenzyl guanidine), a guanethidine analog. The [131 I]MIBG scan has replaced the radionuclide scan as a clinical tool to detect neural crest-derived tumors in opsoclonus-myclonus because it is much more sensitive (293). Circulating markers found in blood or urine include

Disorder	Reference		
Amaurosis congenita of Leber ^A	264		
Congenital/neonatal ^A	76, 263, 264		
Vertebrobasilar disorders ^{A,B}	265, 266		
Tectocerebellar dysraphia with occipital encephalocele ^A	267		
Multiple sclerosis ^A	71, 268		
Friedreich's ataxia ^A	71		
Craniofacial dysmorphism ^A	269		
Smith-Lemli-Opitz syndrome ^A	270		
Familial cerebellar vermis atrophy ^A	271		
Thalamic hemorrhage ^A	272		
Pontine hemorrhage	102		
Hyperosmolar nonketotic coma ^{A,B}	52, 95, 273		
Demyelinating disease ^A	2		
Multiple carboxylase deficiency ^{A,B}	262		
Hydrocephalus ^A	81		
Lafora disease ^{A,B}	272		
Head trauma ^{A,B}	275, 276		
Palatal myoclonus ^{A,B}	275, 277, 278, 279, 280		

TABLE 7. Other disorders associated with $opsoclonus^A$ or $myoclonus^B$

neuron-specific enolase, ferritin, and gangliosides (294). Elevated plasma and tumor concentrations of the disialoganglioside G_{D2} is detected in children with undifferentiated neuroblastoma but not with ganglioneuroblastoma or ganglioneuroma. Tumors lacking G_{T1b} may signify a poor prognosis. Mediastinal neuroblastomas contain more complex b-series gangliosides (G_{D1b} and G_{T1b}) than monosialogangliosides, indicating a more differentiated cellular or membrane composition.

Endogenous and exogenous factors determine the diferentiation and proliferation of neuroblastomas. Endogenous factors include patient age, biological tumor maturation, tumor site innervation, proto-oncogenes, cellular receptors, and peptide growth factors (295). Nerve growth factor (NGF) exerts both differentiating and mitogenic effects on neural structures. Little use of these various tumor markers has been made in opsoclonus-myoclonus.

Clinical and Molecular Genetics

Neuroblastomas have occurred in siblings (296,297), and familial cases often involve multiple primary tumors and are diagnosed at younger ages than sporadic cases. Benign and malignant tumors of neural crest origin occur with increased frequency in neurofibromatosis, an autosomal-dominant disorder. Host or genetic factors are also supported by the hereditary association of essential myoclonus (not opsoclonus) and malignant melanoma (298). A small group of neuroblastomas are associated with deletion of chromosome 13, and may be accompanied by congenital malformations. These observations led to the "two-hit hypothesis" of neuroblastoma, i.e., at least two gene abnormalities are requisite for tumorigenesis, allowing for an inherited factor and some other factor (299). The hypotheses of pathogenesis in paraneoplastic opsoclonus-myoclonus (20,27,29,159,299) are as follows:

Primary carcinogens, simultaneous but independent brain damage and tumor induction. Extrinsic carcinogen may be chemical or virus

Brain damage from metabolite (catecholamine?) liberated by tumor Immune-complex disease

Hereditary factors in genesis of tumor and neural damage with extrinsic carcinogen

Chromosomal deletions and DNA amplifications have been found in human neuroblastoma. The deleted genes (short arm of chromosome 1) may be tumorsuppressor genes, whereas the amplified genes (multiple gene copies) are cellular oncogenes (300). DMs (double minutes) and HSRs (homogeneously staining chromosome regions) are the sites of amplification of N-myc (chromosome 2p). Amplified L-myc (chromosome 1p) genes are found in human small cell lung cancer cells. N-myc is highly expressed in undifferentiated neuroblasts but not in differentiated ganglion cells. The clinical correlate of N-myc expression appears to be advanced tumor stage (Evans III–IV) (206) and worsened prognosis. N-myc is also amplified in normal fetal brain. Significant aneuploidy (near triploid) in the absence of chromosome 1p deletions and N-myc amplification carries a good prognosis in contrast to near diploidy (301).

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Opsoclonus-myoclonus has been reported in second cousins (24). Opsoclonusmyoclonus with neuroblastoma has been reported in Turner syndrome and hemoglobin SC disease (302,303). In four neuroblastomas from children with opsoclonus-myoclonus, single copies of the N-myc oncogene have been found (304).

Monoamines

Catecholamines are often secreted by neuroblastomas and are detected in the urine as the metabolites VMA and HVA in 47–95% of the cases (305–308). The methionine metabolite, cystathionine, is present in urine in 50% of children with neuroblastoma but not in normal children (309).

Could excessive tumor-secreted catecholamines or metabolites induce opsoclonus-myoclonus? It seems unlikely. Most opsoclonus-myoclonus cases with neural crest tumors are not associated with increased urinary catecholamines (4,310, 311), and when they are, tumor removal returns circulating catecholamine levels to normal, but opsoclonus-myoclonus may persist or return (5). Catecholamine neurotransmitters are not lipid-soluble and do not cross the blood-brain barrier. Most hormone-secreting neural crest tumors are not associated with opsoclonusmyoclonus (27). Pheochromocytomas secrete the same catecholamines (other chemicals also), but are not associated with opsoclonus-myoclonus or other dyskinesias.

Tumor catecholamines have been measured as well (312). They do not correlate well with urinary catecholamine excretion patterns or with the degree of histologic differentiation, as seen by light microscopy. A more favorable clinical stage (I, II, IV-S), age more than 1 year at diagnosis, and survival are associated with tumors exhibiting more differentiated patterns of catecholamine metabolism (206). In these tumors, norepinephrine is increased relative to dopamine and dopa (as urine normetanephrine and VMA are increased relative to dopamine and HVA). Serotonin uptake also has been reported in cultured neuroblastoma cells (313).

Immunologic Tests

Several lines of evidence support an immune mechanism of paraneoplastic and parainfectious opsoclonus-myoclonus (20,29,49,314–319) as follows:

Natural history of spontaneous regression of neuroblastoma

Lymphocytic infiltrates in tumors from patients with good prognosis

Lymphocytes cytotoxic to neuroblastoma from affected patients

Anti-neurofilament antibodies

Neurologic improvement in some patients after tumor resection or chemotherapy

Response to ACTH or steroids-immunosuppressive?

Quantitative serum IgG abnormalities with CSF plasmacytosis

Better prognosis for survival of patients with paraneoplastic syndrome

(implies enhanced autoimmunity which controls tumor growth and spread)

The immune-mediated disorder, myasthenia gravis has been a presentation of neuroblastoma (320). The author is aware of a child with both paraneoplastic opsoclonus-myoclonus and myasthenia gravis.

Circulating anti-neurofilament protein antibodies (MW 210K) were found in sera from two children with opsoclonus-myoclonus (no tumor disclosed) using the immunoblot technique (49). The antibodies, which were not found in CSF, disappeared during treatment with adrenocorticotropic hormone (ACTH) or steroids, when clinical symptoms were alleviated. One of the cases also studied by immunofluorescence exhibited antibodies (IgG) which weakly stained neurofibrillary and membrane components of guinea pig Purkinje cells and rat peripheral nerve axons (321). His immunofluorescence titers fell 5 weeks after beginning ACTH therapy. Sera from 21 children with other neurologic disorders did not stain any proteins in the brain homogenate. Neurofilaments are crucial to the developing nervous system; however, neurofilament protein antibodies have been found in sera from several degenerative neurologic disorders as well as normals (322), and may therefore lack specificity.

In six children with opsoclonus-myoclonus, circulating cerebellar-specific immunoreactivity (MW 27K, 35K, and 62K) was found (323).

No anti-CNS antibodies were found in an adult patient with opsoclonus with ataxia and oat cell carcinoma of the lung (129). Serum antibodies to human Purkinje cells were present in a woman with Stage I intraductal breast cancer, and tumor RNA contained the message coding for paraneoplastic cerebellar degeneration (PCD)-related protein (200,324). In several cases of PCD, circulating antibodies to Purkinje cells have been found (325–329). A few of the patients improved after tumor resection or treatment, which correlated with a fall in antibody titer in one case (330).

In adults, certain autoantibodies suggest a paraneoplastic syndrome due to a specific tumor. Anti-Yo is associated with cerebellar degeneration and gyneco-logical cancers (325), Anti-Hu with small cell lung cancer (331–333), and Anti-RI with opsoclonus and breast cancer (334).

The absence of an autoantibody does not rule out a paraneoplastic syndrome, since many patients do not harbor measurable autoantibodies. This appears to be particularly true for children with opsoclonus-myoclonus who do not exhibit any of the antibodies found in the adult paraneoplastic syndromes (334). It should be noted that the cerebellum is the brain region used to test for autoantibodies. Whereas the rationale for this is strong in adult paraneoplastic cases where cerebellar pathology has been found, it is weak in its application to pediatric cases. There have been no studies using brainstem, which may be more appropriate.

More than 50 monoclonal antibodies have been reported to bind to neuroblastoma cells, but no truly neuroblastoma-specific monoclonal antibody has been found (335). A panel of highly specific monoclonal antibodies is required to diagnose neuroblastoma. Neuroectodermally derived cells possess common antigenic determinants or molecules on the cell surface membrane. Autoantibodies to neuroblastoma cell surface antigens have been reported in neuropsychiatric lupus (336). Neuroblastoma cells contain immunoreactive neurofilament proteins (337). Cross-antigenic reactions between T-cell subsets and Purkinje cells have been reported (338).

Leukocytes from children with opsoclonus showed an abnormal reactivity to neuroblastoma extract (32). Suppressor T-lymphocyte function was depressed in

an adult patient with opsoclonus and breast carcinoma, and the abnormality was reversed with prednisone in parallel with improvement of opsoclonus (339).

Circulating autoantibodies to ACTH were found in one of six children with opsoclonus-myoclonus studied, who had been treated with ACTH for several years with loss of efficacy (340). Lymphocytic infiltrate was noted in the tumors of several patients with opsoclonus-myoclonus (134,302).

TREATMENT

ACTH and Steroids

Symptomatic initial responsiveness to ACTH in children (40 IU per day) may occur in 80–90% of cases (Table 8) (12,20,27,29,65,103). ACTH trials have not been reported in adults. Initial response to ACTH in a few days and complete resolution of opsoclonus and myoclonus in 2 weeks has been reported (12,117) (tumor had been resected). Response to ACTH rather than steroids has been described; however, there have been no controlled comparative studies of ACTH versus steroids (12,20,45,46,103,113). One patient completely responded to a single injection of ACTH (20 units) (12). Apparent ACTH dependence may occur with return of symptoms when the dose of ACTH is lowered. The threshold dose is not absolute (25 IU QOD). Some patients with opsoclonus-myoclonus have remained on ACTH or steroids for years (11,340). Side effects from ACTH include cushingoid appearance, cardiovascular abnormalities, hyperpigmentation, and slowed growth (310). ACTH therapy may result in a false-positive gallium scan (107). Not all patients respond to either ACTH or steroids (5,45) and one case apparently worsened (5).

ACTH₁₋₂₄ (208,276,341) and ACTH₁₋₃₉ (Acthar) (3,12,120) have been used successfully, but no trials of ACTH fragments have been reported in opsoclonusmyoclonus. Dexamethasone (11,45,86,112), prednisone (100,102,107,112,117, 287), prednisolone (7,19), triamcinolone (11), β -methasone (18), hydrocortisone (20), and unspecified steroids have been used successfully. Rapid response to steroids (within 3 days) (45) and steroid dependence (11,45) have been reported.

Response to ACTH or steroids does not differentiate patients with and without tumors (65), and therefore should not preclude need for extensive diagnostic evaluation. There are no data regarding whether early treatment with ACTH or steroids masks or lessens the chances of being able to diagnose an underlying neoplasm. In one infant, ACTH treatment failed and subsequently tumor was found (3). ACTH and steroids may be only symptomatic therapies since long-term outcome does not correlate with drug response (65,137,139). One author reserved prednisone for paraneoplastic cases resistant to cyclophosphamide (342).

It may be noteworthy that an adult patient with metastatic medullary carcinomina of the thyroid (and multiple endocrine neoplasia type I) developed opsoclonus-myoclonus after adrenalectomy for ectopic production of ACTH causing Cushing's syndrome (83).

Drug Therapy

Several other drugs have been used in opsoclonus-myoclonus syndromes (19,23,45,46,62,65,84,88,91,103,104,111,128,131,133,134,137,142,261,282,287,315,

Drug	Reference		
Acetylcholine			
Trihexiphenidyl	46, 19		
ACTH			
ACTH ₁₋₃₉	3, 12, 120		
ACTH ₁₋₂₄	209, 276, 341		
Adrenergics	, ,		
Propranolol	23, 45, 65		
Anticonvulsants			
Carbamazepine	19, 128		
Diphenylhydantoin	3		
Phenobarbital	3, 102		
Primidone	128		
Diazepam	3, 19, 60, 91, 103		
Clonazepam	45, 62, 84, 114, 134, 141, 282, 343, 384		
	26, 345		
Nitrazepam	3		
Paraldehyde	128		
Sodium valproate	128		
Chlorazepate	142		
Antihistaminics	2		
Diphenhydramine	3		
Meclizine	3		
Calcium channel blockers	120		
Cinnarizine	128		
Depleters			
Reserpine	111		
Dopamine			
L-dopa	91		
Bromocriptine	91		
Lisuride	62		
Prochlorperazine	128, 287		
Promethazine	46		
Haloperidol	19, 91		
GABA			
Baclofen	62, 133, 287		
Other			
Thiamine	62, 83		
Piracetam	62		
Azathioprine	134		
Biotin	262		
Nialamide	45		
Serotonin			
L-5-HTP	89, 104		
Periactin	46		
	46, 62		
Methysergide Staroida	TU, UL		
Steroids	11, 45, 85		
Dexamethasone			
Prednisone	100, 101, 107, 117, 112		
Prednisolone	7		
Triamcinolone	11		
β-Methasone	18		
Hydrocortisone	20		

 TABLE 8. Attempted pharmacologic therapies

343–345) (Table 8), but there have been no large or controlled studies. Anticonvulsants, including diphenylhydantoin (3,11), phenobarbital and diazepam (3,12), and paraldehyde are not effective, but thiopental abolished opsoclonus and myoclonus intraoperatively (40). Often, the anticonvulsant was not specified or drugs

were used in rapid succession without washout. Patients infrequently respond to clonazepam (84); but response to clonazepam was reported in two patients who failed to respond to corticosteroids and propranolol (282,343). The results of propranolol have been mixed: No response (45,65,282) versus improvement (23) (the two responders had not improved with steroids). Other drugs used unsuccessfully in a few cases include haloperidol, L-dopa, bromocriptine, baclofen, meclizine, promethazine, prochlorperazine, trihexyphenidyl, and cyproheptadine. Improvement of myoclonus was reported following intravenous 5-hydroxy-L-tryptophan (L-5-HTP) in one steroid-dependent child (88) and in two adults (104). However, opsoclonus responded in the adults but not the children. Myoclonus responded to thyrotropin-releasing hormone (TRH) or a TRH analog in the same two adult patients (104). Thiamine-responsive opsoclonus was reported in a case of bronchogenic carcinoma (56) but other patients with paraneoplastic opsoclonus have not responded to thiamine (287). Biotin-responsive opsoclonusmyoclonus was reported in a case of multiple carboxylase deficiency (262). The novel antimyoclonic drug piracetam, which is effective in half the cases of cortical myoclonus of other etiologies but in no cases of subcortical myoclonus (346), was not effective in one adult with paraneoplastic opsoclonus-myoclonus (62). The pharmacologic treatment of myoclonus has been reviewed elsewhere (347).

Plasmaphoresis

Plasmaphoresis has been used unsuccessfully in a few cases of paraneoplastic opsoclonus-myoclonus in adults (62,136).

Tumor Therapy

Tumor removal may permanently decrease opsoclonus and myoclonus (3,59, 67,73,103,175), have a partial effect (102), no effect (172), or cause exacerbation. Slow spontaneous resolution of neurologic symptoms may take months or years (5,29). In one case, institution of radiation therapy appeared to be associated with exacerbation (103). As many of half of the cases were found to show neurologic improvement within one month following surgical excision of neuroblastoma (65).

Chemotherapy, which is also immunotherapy, without surgical resection or radiation, although effective tumor therapy, has induced neurologic remission in a few (46,62). Chemotherapy has included cyclophosphamide (45,58), vincristine (302), both (7), plus other drugs (29). More often, chemotherapy, surgical, and radiation therapies are combined (58). The tumor usually does not recur (4,26,27, 31,58,59,114,302), neurologic remission sometimes occurs (7) but is not the rule (29,38,45,46,302). In cases with previously elevated urinary VMA levels, treatment of the tumor may normalize the levels within days (103). In one case, persistently elevated levels of VMA in the urine despite surgery, radiation, and chemotherapy prompted a second laparotomy, with the finding of tumor recurrence (7).

γ-Globulin

Recently, several patients have been given γ -globulin injections empirically. Word-of-mouth reviews are mixed. There are no published data.

NEUROLOGIC OUTCOME

Neurologic outcome does not appear to depend on etiology, age of onset, early treatment, or relapses after infection. Comments on tumors and treatments have already been made.

There is a paucity of data on the clinical features of the chronic syndrome. Myoclonus, opsoclonus, ataxia, and cognitive problems may each respond differently to therapy (11), and cognitive problems may persist in the apparent absence of motor problems (26,29,31,58). The most common chronic motor abnormality is ataxia (16,134).

The nature of cognitive abnormalities is less clear. Several authors use the term mental or psychomotor retardation which occurs in 61 to 82% of the cases regardless of etiology (5,16). Exact results of neuropsychiatric testing are seldom given. IQs of 58–66 have been documented in a few cases (63): Subscores were not provided, but trouble with language was noted. An IQ of 105 was reported in another child with complete recovery (35). One four-year-old child scored only 50 on the Stanford-Binet test.

In one study, 7 of 10 patients had deficits in cognition or intellect, hyperactivity, impulsivity, or emotional lability (284). Hyperactivity, poor speech, and short attention span were found in a few children with severe intellectual impairment (11,29,302). In another report, in 6 of 26 cases not found to be mentally retarded, significant educational handicaps were found (16).

Fatalities are uncommon, but have been reported even for opsoclonusmyoclonus without tumor (16). There are no other reports of a shortened life expectancy. Long-term outcome studies in opsoclonus-myoclonus are needed.

PHARMACOLOGIC HYPOTHESES OF PATHOGENESIS

While altered neurotransmission is the basis of myoclonus (347), there is little direct information linking opsoclonus-myoclonus to a specific neurotransmitter or abnormality. Several lines of evidence, however, suggest a pharmacologic model, although the data are still insufficient to construct one (348).

Molecular Action of ACTH

ACTH may exert its antimyoclonic action by its effects on the brain, functioning as a neurotransmitter or modulating the activity of neurotransmitters. Focus on the brain is appealing because it would explain how ACTH could be effective treatment despite diverse etiologies of opsoclonus-myoclonus.

ACTH may alter neurotransmission in several ways. The redundancy of behavioral information in ACTH peptides is consistent with a multiplicity of binding sites (of high affinity and low capacity) in brain (349). Micromolar activity of ACTH₁₋₃₉ or ACTH₁₋₂₄ has been reported in vitro at serotonin (5-HT) (350), *N*-methyl-D-aspartate (NMDA) (351), opiate and dopamine (352–355), but not benzodiazepine receptors (356). It is interesting that, at all neurotransmitter receptors for which ACTH₁₋₃₉ and ACTH₁₋₂₄ have shown activity in vitro, the

nonsteroidal fragments $ACTH_{4-9}$ or $ACTH_{4-10}$ have been less active or inactive (357).

Only the long ACTH fragments potently increase cyclic-AMP, stimulate phosphoinositide hydrolysis and inhibit protein kinase phosphorylation, enhance lipid fluidity of synaptic plasma membranes, and stimulate protein synthesis (358–359). Besides weak effects at classic neurotransmitter receptor sites, ACTH also increases dendritic arborization in developing brain (360) and exercises other neuromodulatory effects on neurotransmission (349,361,362). The significance of micromolar receptor effects of ACTH is uncertain in view of all the different effects of ACTH which may contribute to its clinical properties. Peptides, because of their tertiary conformation, may interact nonspecifically with a variety of receptors. Although trophic effects on receptor density with chronic ACTH administration have been reported (363–366), the rapid effect of ACTH clinically may require action as a neurotransmitter rather than a neuromodulator. ACTH is found in brainstem in theoretical proximity to "myoclonic centers" (367–368) and its secretion is under monoaminergic regulation in man (369).

Glucocorticoids, which are a consequence of ACTH treatment, stimulate adrenergic differentiation of neural crest tumor cells in culture (370). Although ACTH may modify paraneoplastic opsoclonus-myoclonus through an effect directed at tumor, not brain, this mechanism would not explain its efficacy in opsoclonus-myoclonus due to other etiologies.

Tumor Receptors

If one accepts the assumption that the underlying mechanism of opsoclonusmyoclonus is immunologic (29), then it is necessary to identify the antigenic stimulus shared by tumor and brain. There are many possible candidate substances, but a lipoprotein surface-membrane antigen, one which is involved in neurotransmission and the pharmacology of myoclonus, would appear logical. There is evidence that both host and tumor factors participate in the pathogenesis of this paraneoplastic syndrome. The neurobiological problem is that the mechanism by which only the minority of peripherally located neural crest tumors induce brain dysfunction has remained both intriguing and elusive. Neural crest tumors which induce opsoclonus-myoclonus may be biologically different, since patient survival is greater, the tumor is histologically more differentiated, levels of circulating catecholamines are lower, and the tumor is more often a ganglioneuroblastoma and of thoracic location (46,97,109,312). It is possible that excessive monoamines could be immunogenic, since autoantibody formation associated with methyldopa therapy has been reported (371).

The hypothesis that brain neurotransmitter receptors are the target of autoantibodies to neural tumor receptors or receptor-active tumor products is novel (348). This hypothesis is supported by the identification of neurotransmitter receptors relevant to opsoclonus and myoclonus, such as serotonin, adrenergic, opiate, and cholinergic receptors, in rodent neuroblastoma hybrid cell lines (372–382), research development of monoclonal antibodies to neurotransmitter receptors (383– 385), the finding of antibodies in opsoclonus-myoclonus (49), and evidence that antibodies to neurotransmitter receptors cause neurologic disease (386–388) by autoantibody destruction of neurotransmitter receptors or intrinsic biological activity at those receptors (389). In the central nervous system, autoantibodies directed against GABAergic neurons have been found in stiff-man syndrome (390). Cross-antigenic reactions have also been found between natural killer cells and nervous tissue and between subgroups of T cells and Purkinje cells (338,391).

Human neural crest-derived tumors contain different populations of neurotransmitter receptors. The serotonin 5-HT_{1A}-like receptor recognition site may be a new biologic marker differentiating human ganglioneuroblastomas from neural crest-derived tumors (392-393). The finding that 5-HT_{1A}-like sites are expressed by ganglioneuroblastoma, the tumor most often associated with pediatric opsoclonus-myoclonus, but not by neuroblastoma or ganglioneuroma (394), is highly relevant to animal studies linking myoclonus to 5-HT_{1A} receptors (395). 5-HT₃ and 5-HT_{1E} binding sites have been found in human neuroblastomas (396). Tumor serotonin receptors may be relevant because L-5-HTP is useful in some forms of myoclonus, particularly posthypoxic action myoclonus (397,398). Its therapeutic action is dependent on decarboxylation to 5-HT in the CNS. 5-HT has equal affinity for the various 5-HT₁ receptors. Chronic treatment with L-5-HTP in the rat alters cortical 5-HT₂ receptors (399). Delineation of the role of each of these subtypes in human myoclonus and in the action of L-5-HTP is incomplete. In rodents, involvement of the 5-HT_{1A} and the functionally linked 5-HT₂ receptor has been identified (395). However, the other newer sites have not been studied. Antibodies to 5-HT_{1A} receptors have been reported in CSF of some patients with autism (400). However, other neural crest tumors beside ganglioneuroblastoma evoke opsoclonus-myoclonus.

Receptors must be measured in tumors which evoke the paraneoplastic syndrome to test this hypothesis. Further, without a complete survey of neurotransmitter receptors on human neural crest-derived tumors, it would be premature to focus on any one receptor. Some human neural crest tumors also contain peripheral benzodiazepine receptors (401). Nerve growth factor receptors are found both in neuroblastoma and cerebellar Purkinje cells. Receptors for NGF are located on the cell-surface membrane, neuronal synaptic terminals, and the nucleus. A "slow" NGF receptor on the plasma membrane is the probable NGF binding site for biological responses such as neurite outgrowth.

Cell surface receptors on neural crest-derived tumors have implications both for oncogenesis and chemotherapy. 5-HT receptors and perhaps other tumor receptors, which mediate the effect of 5-HT as a growth and differentiating factor in developing brain, may trigger malignant transformation; the 5-HT receptor functions as a protooncogene (402). In contrast, tumor receptors may also be a target for chemotherapy. The neurotoxic analogue of dopamine, 6-hydroxydopamine, enters neuroblastoma cells via surface catecholamine receptors, and reduces tumor growth (403).

The p,p'-DDT Myoclonic Model

Since the insecticide DDT induces opsoclonus in humans and myoclonus in both humans and animals (404,405), it may be useful to review the involvement of

neurotransmitters in its action. The cerebellum may be the principal site of action of DDT (405). In the p,p'-DDT mouse model, L-5-HTP is antimyoclonic (406). Microinjection of p,p'-DDT into the inferior olive, cerebellar dentate nucleus, or the red nucleus (280,407) (Guillain-Mollaret triangle) induces myoclonus in the rat.

A related insecticide, chlordecone, increases 5-HT turnover, reduces the density of 5-HT₁ but not 5-HT₂ receptors in striatum and hippocampus, and induces tremors blocked by 5-HT antagonists (408).

A new, potential antimyoclonic therapy is the glycine prodrug milacemide (2*n*-pentylaminoacetamide). Milacemide, an effective anticonvulsant, readily penetrates to the brain and is metabolized primarily to glycine and glycinamide (409). γ -Aminobutyric acid (GABA) levels are also increased in the basal ganglia. The therapeutic index of milacemide is high (410). In the *p*,*p'*-DDT model, milacemide is antimyoclonic (411). Monoamine oxidases participate in the deamination of milacemide to glycinamide.

REM Sleep Myoclonus and Atonia

Sporadic myoclonus occurs physiologically, although paradoxically, during the rapid eye movement (REM) periods of active sleep (412) when the body is otherwise functionally paralyzed and flaccid (atonia) (413). Sleep also regulates eye movements (414,415). This is a dynamic process which is actively regulated, but may fail. Atonia may be pathologically absent during REM sleep (416). Because pathologic myoclonus and opsoclonus may persist during sleep, the neural mechanisms of REM motor control also may be dysfunctional in the opsoclonus-myoclonus syndrome. The anatomic and pharmacologic basis of those mechanisms may shed light on the pathophysiology of the opsoclonus-myoclonus syndrome. The classic view is that myoclonus is evoked by sensory influxes at the medullary level when forebrain inhibition is lost (417).

Myoclonus due to lesions of the lower brainstem or spinal cord increases during light non-REM (NREM) sleep, but attenuates during REM (418). Activation of myoclonus under these circumstances may be due to dissociation of spinal α - and γ -motoneuronal activity. In NREM, α -motoneuron activity is depressed but γ -activity is unchanged, whereas in REM, the activity of both motoneurons is depressed (419). In contrast, opsoclonus-myoclonus is diminished in light NREM (Stage 2) and reappears in REM (86). This same pattern applies to the ocular oscillations and myoclonus of palato-ocular myoclonus which results from various intrinsic brainstem lesions (66).

REM periods are characterized by increased motoneuron inhibition which results in atonia, and bursts of overpowering excitation, which result in myoclonus. The inhibition [large amplitude inhibitory postsynaptic potentials (IPSPs)] is apparently mediated by glycine. The excitations [excitatory postsynaptic potentials (EPSPs)], which are mediated by a non-NMDA neurotransmitter (420), begin as hyperpolarization but are followed rapidly by a depolarizing shift and an action potential (420).

Myoclonus may reflect the high activity of the nervous system during REM

sleep, when cortical and subcortical neurons discharge at uncharacteristically rapid rates and activity along motor pathways is enhanced (421).

Both tonic and phasic aspects of REM sleep result from brainstem activity (422). Deinhibitory drive originates in the pons with a cholinoceptive trigger zone in the dorsolateral pontine tegmentum, which may be the nucleus pontis oralis (420,423) or the peri-locus ceruleus alpha (424). The excitatory drive emanates from the medulla in the nucleus gigantocellularis reticularis (420,423) or the adjacent nucleus reticularis magnocellularis (424). The nucleus gigantocellularis reticularis is known from previous studies to be a myoclonus generator (417). Microinjection of NMDA agonists into the nucleus magnocellularis of the decerebrate cat induces myoclonus (and increased muscle tone) and NMDA antagonists block the myoclonus (but not the muscle tone) (425). Of possible relevance to opsoclonus-myoclonus is the observation that corticotropin-releasing factor (CRF), which releases ACTH in brain, also inhibits NMDA agonist-induced myoclonus.

The Saccadic System

The saccadic system depends on the interaction of "burst" cells and "omnipause" cells both of which reside in the brainstem (426). Burst neurons, located in the paramedian pontine reticular formation, are silent until just before or during a saccade, when they drive ocular motor neurons to create saccades (427). In contrast, omnipause neurons cease firing when burst cells fire and inhibit burst cells during fixation (426).

The mechanism of opsoclonus or ocular flutter have been attributed to a disorder of burst cells (428) or in pause cell control of burst neurons (427). Increased saccadic velocities with normal amplitude (428) would favor excessive burst cell discharge, but have not been uniformly recorded (429). The presence of this "brainstem generator" for saccades suggests a brainstem origin for opsoclonus and a cerebellar locus for ocular flutter and dysmetria (2,272). There is no consensus, however. The mesencephalic tegmentum (171) has been implicated in opsoclonus. The continuum of opsoclonus, ocular flutter, and ocular dysmetria has also been related to cerebellar disease (429). The influence of the cerebellum on eye movements (430,431) has led some to argue that opsoclonus is purely cerebellar (432). Similarly, myoclonus has been attributed to the cerebellum (433,434), but such cases are more likely due to a brainstem mechanism for myoclonus possibly with cerebellar inputs which the cerebellar lesion has disrupted. Supratentorial mechanisms also influence saccades (435,436).

Putative omnipause neurons in two patients with opsoclonus associated with oat cell lung carcinoma were normal by light microscopy (437). However, the location of the respective neurons in humans has not been established. In an idiopathic case of opsoclonus-myoclonus, no abnormalities were seen in the paramedian pontine reticular formation of the caudal pons (438).

The neurotransmitters for burst cells and omnipause cells are not known. Methyltyrosine (439) and L-tryptophan (440) produce square-wave jerks in normals implicating monoamines in pause cell control (439). Saccades can be modified experimentally by GABA-ergic drugs (441).

Neuropathologic Clues

In one autopsied case, brain changes were restricted to the cerebellum and included peridentatal demyelination, gliosis, and loss of Purkinje cells (110).

In adult paraneoplastic syndromes, variable histologic abnormalities have been described including mild Purkinje cell loss, lymphocyte infiltration, cerebellar gliosis, edema, and demyelination around the cerebellar dentate nucleus (99,174, 442).

Pathologic changes in patients with clinical brainstem involvement have been found most often in the medulla or pons, especially in the inferior olivary nuclei (168,443–445). A so-called idiopathic case had both cerebellar and inferior olivary lesions (438). Findings suggestive of midbrain encephalitis have been described as a remote effect of malignant neoplasm (446).

A child with opsoclonus-myoclonus was biopsied because neuroradiological studies indicated a lesion in the cerebellar vermis: Purkinje and granular cell loss with gliosis were found (288). Pathologic changes of encephalomyelitis restricted to lower medulla and upper cervical spinal cord were found in an adult with opsoclonus-associated oat cell lung carcinoma (287). Loss of internuncial spinal neurons was found in a case of opsoclonus with myoclonus but the brainstem was not studied (447). A frontal cortical biopsy in another child revealed no significant histological abnormalities (12).

It has been suggested on the basis of stroke anatomy in the vertebrobasilar circulation in four patients that opsoclonus with palatal myoclonus indicates lesions of the upper cerebellar crus (path of the dentato-olivary fibers and fibers connecting the cerebellar flocculus with oculomotor nuclei) (279). Palatal myoclonus alone suggests lesions instead of the cerebellar dentate nucleus and central tegmental tract of the brainstem. There was a delay of 1.5 to 4 months before opsoclonus and/or myoclonus appeared. Opsoclonus (with myoclonus) may occur with ocular bobbing, which is usually pontine (448).

Cognitive Functions of Cerebellum and Motor Nuclei

It has been suggested that the occurrence of mental retardation in opsoclonusmyoclonus indicates a more widespread effect on the CNS than on the cerebellum alone (7). However, in animals, there is evidence that the cerebellum and inferior olives influence learning (449). It has been proposed that the cerebellum is the seat of motor learning through the mechanism of climbing fiber synapses on Purkinje cells (450). However, lesions of the inferior olives, from which climbing fibers emanate, prevent motor learning, suggesting that the inferior olives are principally involved (451). The cerebellum contributes to oculomotor plasticity. There is some evidence that the cerebellum also has a role in cognitive behaviors besides motor learning, for which the repository site of learning may be the cerebellar target areas rather than the cerebellum itself (449,452). These behaviors include spatial learning (connections between cerebellum and frontoparietal association cortex, limbic system, and superior colliculus), discrimination learning (projections to cerebellum via pontine nuclei), and emotions such as fear (reticular activating system, limbic system, and hypothalamus) (449). In adults with psychiatric disorders and cerebellar lesions, cerebellar dysfunction may contribute to a syndrome of reduced memory, concentration, abstraction, labile affect, and impaired social skills and environmental adaptation (453). In children, cerebellar pathology has been described in autism (454).

CONCLUSIONS

Myoclonus in opsoclonus-myoclonus is predominantly evoked by action and sensory stimuli and often results in major functional impairment of gait and sitting. The tremulous appearance of these patients is more likely due to the characteristically small amplitude and rapid myoclonic jerks than to the presence of tremor. The relative contribution of cerebellar dysfunction in opsoclonus-myoclonus is unclear because myoclonus-induced motor incoordination may be difficult to differentiate from primary cerebellar ataxia. Both may contribute to the dysarthria of some patients. While the syndrome may resolve, permanent neuropsychiatric sequelae are common in children, and myoclonus has less of a place in the chronic phase than ataxia and learning and behavioral problems. The occurrence of anxiety is so common in the acute phase that hypothesizing an associated anxiety disorder may contribute to understanding the pathophysiology.

Opsoclonus-myoclonus is one of the few recognized paraneoplastic dyskinesias; it is a remote effect of certain tumors. As a putative autoimmune-mediated dyskinesia, it is part of a group that includes Sydenham's chorea, and chorea of CNS-lupus. It also overlaps with the syndrome of acute cerebellar ataxia of childhood. Opsoclonus-myoclonus is a syndrome, not a single disease, which also may be induced by drugs and toxins, metabolic disorders, congenital and degenerative disorders, and infections. An immunologic theory of opsoclonus-myoclonus may explain its association with a viral syndrome or with peripheral tumors. The tumors in children tend to be histologically and biochemically more differentiated, more often thoracic, and associated with a greater survival rate than those without a paraneoplastic syndrome. Molecular genetic studies of cellular oncogenes have not identified a difference between tumors with and without a paraneoplastic syndrome, despite the chromosomal and DNA abnormalities found in human neuroblastoma. Various circulating anti-CNS antibodies have been found in paraneoplastic syndromes in adults, but only an anti-neurofilament protein antibody has been detected in pediatric opsoclonus-myoclonus. The absence in children with opsoclonus-myoclonus of the autoantibodies typically found in adults may be due to using the wrong brain region against which to test for antibodies: cerebellum rather than brainstem.

Understanding the locus of opsoclonus-myoclonus may rely more on theoretical clues than from neuropathologic or neuroradiologic studies, which are unrevealing in the majority of cases. The proximal and distal generalized distribution of myoclonus, absence of enlarged somatosensory evoked potentials, and rarity of associated seizures favors a subcortical origin of myoclonus. Opsoclonus has only slightly greater localizing value than myoclonus, but a primary brainstem mechanism for each is likely with regulatory inputs from cerebellum and cerebrum. The mechanism for opsoclonus and myoclonus are independent but neighboring since either disorder may occur separately but their co-occurrence is not random. If the occurrence of opsoclonus-myoclonus were indicative of a diffuse injury, convulsions, other dyskinesias besides myoclonus, paresis, and coma should be more prevalent; but in children with opsoclonus-myoclonus, they are rare. Therefore, the injury of subcortical structures with dual motor and cognitive functions, such as the inferior olive and cerebellum, is plausible (this may be analogous to the occurrence of obsessive-compulsive disorders in Sydenham's chorea). A brainstem origin is supported by the location of burst and omnipause cells of the primate saccadic system and the role of the brainstem in both the tonic and phasic aspects of REM sleep, during which myoclonic jerks may appear. Failure of glycine-mediated inhibition or excessive activity of a non-NMDA excitatory neurotransmitter in the medullary reticulum has been proposed.

Little is known about the pharmacology of opsoclonus-myoclonus because neurochemical data are lacking and there have been inadequate clinical drug trials. ACTH is the most often used and effective drug in childhood-onset opsoclonusmyoclonus but is also problematic. The main issue which must be clarified is whether ACTH exerts its antimyoclonic action by suppressing ongoing antibodymediated injury to the CNS or by modifying neurotransmission, as a neurotransmitter or neuromodulator. If ACTH is providing only symptomatic therapy, then it may be supplanted by other more selective and less toxic drugs. Clonazepam, propranolol, or L-5-HTP sometimes may be useful. Biotin- and thiamineresponsive etiologies, though rare, should be ruled out. The induction of opsoclonus-myoclonus by a variety of drugs and chemicals supports a neurotransmitter disturbance as the basis of opsoclonus-myoclonus, but no single neurotransmitter seems to mediate the effects of all the diverse drugs and chemicals which can induce opsoclonus or myoclonus. Neurotransmitter receptors on human tumors may provide the link between pharmacologic and immunologic models of opsoclonus-myoclonus. However, since there is no animal model of paraneoplastic opsoclonus-myoclonus, this remains speculative. The capacity of the insecticide $p_{p'}$ -DDT to induce myoclonus in human and animals may be important because the glycine prodrug, milacemide, is antimyoclonic in the $p_{,p'}$ -DDT animal model. More basic research in opsoclonus-myoclonus is needed before treatment can be improved for these unfortunate patients.

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REFERENCES

^{1.} Orzechowski K, Walichiewicz T. Operated cyst of the cerebellum. Lwowski Tydogn Lek 1913; 8:219-27.

Orzechowski K. De l'ataxie dysmetrique des yeux: Remarques sur l'ataxie des yeux dite myoclonique (opsoclonie, opsochorie). J Psychol Neurol 1927;35:1–18.

- 3. Martin ES, Griffith JG. Myoclonic encephalopathy and neuroblastoma. Report of a case with apparent recovery. Am J Dis Child 1971;122:257–8.
- 4. Leonidas JC, Brill CB, Aron AM. Neuroblastoma presenting with myoclonic encephalopathy. *Radiology* 1972;102:87-8.
- 5. Senelick RC, Bray PF, Lahey E, Van Dyk HJL, Johnson DG. Neuroblastoma and myoclonic encephalopathy: Two cases and a review of the literature. J Pediatr Surg 1973;8:623-32.
- 6. Delalieux C, Ebinger G, Maurus R, Sliwowski H. Myoclonic encephalopathy and neuroblastoma [letter]. N Engl J Med 1975;292:46-7.
- Brandt S, Carlsen N, Glening P, Helweg-Larsen J. Encephalopathia myoclonica infantalis (Kinsbourne) and neuroblastoma in children. A report of three cases. *Dev Med Child Neurol* 1974; 16:286–94.
- Despres P, Herouin C, Seringe P. Encephalopathie myoclonique avec opsoclonies. A propos de deux observations. Ann Pediatr 1968;44:705-13.
- 9. Ritz A, Stover B, Emrich R. Myoklonische enzephalopathie (Kinsbourne-syndrom) und Neuroblastom. [Myoclonic encephalopathy (Kinsbourne's syndrome) and neuroblastoma] In: Doose H, ed. Aktuelle Neuropädiatrie. Stuttgart: Thieme, 1977:64–7.
- Palacio M, Ruiz-Benito MA, Mondragon F, Solorzano C, Tovar JA. Encefalopatia mioclonica infantil con opsoclonus y neuroblastoma (dos observaciones). Am Esp Pediatr 1983;18:118-22.
- 11. Christoff N. Myoclonic encephalopathy of infants. A report of two cases and observations on related disorders. Arch Neurol 1969;21:229-34.
- 12. Kinsbourne M. Myoclonic encephalopathy of infants. J Neurol Neurosurg Psychiatry 1962;25: 271-6.
- Lott I, Kinsbourne M. Myoclonic encephalopathy of infants. In: Fahn S, Marsden CD, van Woert MH, eds. Advances in neurology. Myoclonus, Vol. 43; New York: Raven Press, 1986:127-36.
- Takebe Y, Nomura Y, Higashi T. Myoclonic encephalopathy of infant (Kinsbourne). Report of two cases. *Brain Dev* 1978;10:409-15.
- 15. Azzaroli L. Un caso di encefalopatia mioclonica infantile (M. di Kinsbourne). [A case of infantile myoclonic encephalopathy] *Clin Pediatr Bologna* 1978;6:321–32.
- Marshall PC, Brett EM, Wilson J. Myoclonic encephalopathy of childhood (The dancing eye syndrome): A long-term follow-up study [abstract]. *Neurology* 1978;28:348.
- 17. Lipinski GC. Myoclonal encephalopathy in children. Diagnostic and therapeutic problems. Neuropathol Pol 1975;13:311-17.
- Chiba S, Motoya H, Shinoda M, Nakao T. Myoclonic encephalopathy of infants: A report of two cases of dancing eyes syndrome. *Dev Med Child Neurol* 1970;12:767–71.
- 19. Brune Von G, Hertel G, Gruninger W. Syndrom der tanzenden Augen mit myokloner Ataxie. [Syndrome of dancing eyes with myoclonic ataxia] *Monatsbl Augenheilkd* 1971;158:41-8.
- 20. Dyken P, Kolar O. Dancing eyes, dancing feet: Infantile polymyoclonus. Brain 1968;91:305-20.
- Mejlszenkier JD, Rosman NP, Gilbert JJ. Immunologic studies in infantile polymyoclonia [abstract]. Neurology 1978;28:407.
- 22. Manson JI. Infantile polymyoclonia. Proc Aust Assoc Neurol 1973;9:19-22.
- 23. Fowler GW. Propranolol treatment of infantile polymyoclonia. Neuropädiatrie 1976;7:443-48.
- 24. Robinson GC, Jan JE, Dunn HG. Infantile polymyoclonus: Its occurrence in second cousins. *Clin Genet* 1977;11:53-56.
- 25. McLean DR. Polymyoclonia with opsoclonus. Neurology 1970;20:508-12.
- Forster C, Weinmann H. Symptomatic infantile polymyoclonus. Z Kinderheilkd 1971;111:240– 46.
- 27. Moe PG, Nellhaus G. Infantile polymyoclonia-opsoclonus syndrome and neural crest tumours. *Neurology* 1970;20:756–64.
- Williams TH, House RF Jr, Burgert EO Jr, Lynn HB. Unusual manifestations of neuroblastoma: Chronic diarrhea, polymyoclonia-opsoclonus, and erythrocyte abnormalities. *Cancer* 1972;29: 475–80.
- 29. Bray PF, Ziter FA, Lahey ME, Myers CG. The coincidence of neuroblastoma and acute cerebellar encephalopathy. J Pediatr 1969;75:983-90.
- 30. Jahadi MR, Whitcomb JG, Kini R, High RH. The association of acute cerebellar encephalopathy and neuroblastoma. *Henry Ford Hosp Med J* 1976;24:87–90.
- 31. Malmstrom-Groth A. Cerebellar encephalopathy and neuroblastoma. Eur Neurol 1972;7:95-100.
- 32. Stephenson JBP, Graham-Pole J, Ogg L, Cochran AJ. Reactivity to neuroblastoma extracts in childhood cerebellar encephalopathy ("Dancing eyes" syndrome). *Lancet* 1976;2:975.
- 33. Kompf D, Engelhardt A, Dietrich HJ, Neundorfer B. Acute cerebellar encephalitis in adulthood. Nervenarzt 1985;56:431-39.

- 34. Korobkin M, Clark RE, Palubinskas AJ. Occult neuroblastoma and acute cerebellar ataxia in childhood. *Radiology* 1972;102:151-2.
- 35. Pampiglione G, Maia M. Syndrome of rapid irregular movements of eyes and limbs in childhood. Br Med J 1972;1:469-73.
- 36. Lemerle J, Lemerle M, Aicardi J, Messica C, Schweisguth O. Report of three cases of the association of an oculo-cerebello-myoclonic syndrome with a neuroblastoma. Arch Fr Pediatr 1969;26:547-58.
- Fournier A, Ducoulombier H, Cousin H, Carton MM. Oculo-cerebello-myoclonic syndrome and neuroblastoma. J Sci Med Lille 1972;90:189–97.
- Labre F, Bethenod M, Guibaud P, Mamelle JC, Genoud J. Oculo-cerebello-myo-clonic syndrome and neuroblastoma: Apropos of 2 cases. Arch Fr Pediatr 1972;29:411–20.
- 39. Ford FR. Myoclonic encephalopathy of infants (Kinsbourne). In: Diseases of nervous system in infancy, childhood and adolescence, 5th Ed. Springfield IL: Charles C Thomas, 1966:301-3.
- Burrow FA, Seeman RG. Ketamine and myoclonic encephalopathy of infants (Kinsbourne syndrome). Anesth Analg 1982;61:873-5.
- Kaulfersch W, Urban C, Fritsch G, Maurer G. Myoclonic encephalopathy (Kinsbourne syndrome). Pädiatr Pädol 1984;19:279–85.
- 42. Cogan DG. Opsoclonus, body tremulousness, and benign encephalitis. Arch Ophthalmol 1968; 79:545-51.
- 43. Estrin WJ. The serological diagnosis of St. Louis encephalitis in a patient with the syndrome of opsoclonia, body tremulousness and benign encephalitis. Ann Neurol 1977;1:596-8.
- Baringer JR, Sweeney VP, Winkler GF. An acute syndrome of ocular oscillations and truncal myoclonus. Brain 1968;91:473–80.
- Berg BO, Ablin AR, Wang W, Skoglund R. Encephalopathy associated with occult neuroblastoma. J Neurosurg 1974;41:567-72.
- Altman AJ, Baehner RL. Favourable prognosis for survival in children with coincident opsomyoclonus and neuroblastoma. *Cancer* 1976;37:846–52.
- 47. Labrune B, Vignes B, Grenet P. Syndrome opsomyoclonique chez l'enfant. In: Journées parisiennes de pédiatrie. Paris: Flammarion Médécine-Sciences, 1975:227.
- 48. Mikulowski W. Syndrome rare opso-myo-clonique observé chez un enfant au cours d'une encephalite. [Rare opso-myoclonic syndrome observed in a child during encephalitis] Arch Med Enfants 1926;29:279-84.
- 49. Noetzel M, Cawley LP, James VL, Minard BJ, Agrawal HC. Anti-neurofilament protein antibodies in opsoclonus-myoclonus. J Neuroimmunol 1987;15:137-45.
- 50. Leder RM. The opsoclonus-myoclonus syndrome: A review of the literature. Bull Los Angeles Neurol Soc 1981;46:41-50.
- 51. Kuban KC, Ephros MA, Freeman RL, Laffell LB, Bresnan MJ. Syndrome of opsoclonusmyoclonus caused by Coxsackie B3 infection. Ann Neurol 1983;13:69-71.
- 52. Matsumura K, Sonoh M, Tamoka A, Sakuta M. Syndrome of opsoclonus-myoclonus in hyperosmolar nonketotic coma. Ann Neurol 1985;18:623-4.
- 53. Hattori T, Hirayama K, Imai T, Yamoda T, Kojima S. Positive lesions in opsoclonus-myoclonus syndrome shown by MRI. J Neurol Neurosurg Psychiatry 1988;51:1572-5.
- 54. Leopold HC. Opsoclonus- und Myoklonie-syndrom. [The syndrome of opsoclonus-myoclonus] Fortschr Neurol Psychiatr 1985;53:42–54.
- 55. Rosenberg NL. Hearing loss as an initial symptom of the opsoclonus-myoclonus syndrome. Arch Neurol 1984;41:998-9.
- 56. Nausieda PA, Tunner CM, Weiner WJ. Opsoclonic cerebellopathy—a paraneoplastic syndrome responsive to thiamine. Arch Neurol 1981;38:780–1.
- 57. Bachman DS. Opsoclonic cerebellopathy. Spontaneous remissions [letter]. Arch Neurol 1982; 39:387.
- Solomon GE, Chutorian AM. Opsoclonus and occult neuroblastoma. N Engl J Med 1968;279: 475-7.
- 59. Davidson M, Tolentino V, Sapir S. Opsoclonus and neuroblastoma [letter]. N Engl J Med 1968;279:948.
- 60. Brissaud HE, Beauvais P. Opsoclonus and neuroblastoma [letter]. N Engl J Med 1969;280:1242.
- 61. Mancini CL, Livet MO, Bernard R. Le syndrome ataxie-opsoclonies-myoclonies. Ann Pédiatr 1980;5:269-75.
- 62. Anderson NE, Budde-Steffen C, Rosenblum MK, et al. Opsoclonus, myoclonus, ataxia, and encephalopathy in adults with cancer: A distinct paraneoplastic syndrome. *Medicine* 1988;67: 100–109.

- 63. Pinsard N, Pons-cerdan Cl, Mancini J, Livet MO, Bernard R. The syndrome ataxia-opsocloniesmyoclonus. [Ataxia-myoclonus-opsoclonus syndrome] Sem Hop Paris 1981;57:488-94.
- Talon P, Stoll C. Opso-myoclonus syndrome of infancy. New observations. Review of the literature (110 cases). *Pédiatrie* 1985;40:441-9.
- 65. Boltshauser E, Deonna T, Hirt HR. Myoclonic encephalopathy of infants or "dancing eyes syndrome." *Helv Paediatr Acta* 1979;34:119-33.
- 66. Miller NR. Nystagmus and related ocular motility disorders. In: Walsh and Hoyt's *Clinical neuro-ophthalmology*, 4th Ed, Baltimore: Williams & Wilkins, 1985:892-921.
- Toupet M, de Gramont A, Bacri D, Haguenau M, Pepin B. Le flutter-opsoclonie. A propos de trois observations [Flutter-opsoclonus. report on three cases]. *Rev Neurol (Paris)* 1982;138:219– 39.
- 68. Atkin A, Bender MD. Lightning eye movements (ocular myoclonus). J Neurol Sci 1964;1:2-12.
- 69. Alpert JN, Suga H, Perusquia E. Lightning eye movements. J Neurol Sci 1978;27:71-8.
- 70. Means ED, Brumlik J, Hart C. "Lightning eye movements" and generalized "shuddering" associated with cerebellar ataxia. *Neurology* 1967;17:317.
- 71. Cogan DG. Ocular dysmetria: Flutter-like oscillations of the eyes and opsoclonus. Arch Ophthalmol 1954;51;318–35.
- 72. Prier S, Larmande P, Dairou R, Masson M, Cambier J. Oscillations macro-saccardiques au cours d'un cas d'encéphalopathie myoclonique paranéoplastique. [Macro-saccadic oscillations in a case of paraneoplastic myoclonic encephalopathy]. *Rev Neurol (Paris)* 1979;135:339–46.
- 73. Savino PJ, Glasser JS. Opsoclonus pattern of regression in a child with neuroblastoma. Br J Ophthalmol 1975;59:696-8.
- 74. Cooper JE. Eye movements associated with myoclonus. Am J Ophthalmol 1958;46:205.
- 75. Bowen A, Bergman I. Ultrasonic delineation of left adrenal neuroblastoma in a child with opsoclonus. J Clin Ultrasound 1983;11:31.
- 76. Hoyt CS. Neonatal opsoclonus. J Pediatr Ophthalmol 1977;14:274.
- 77. Smith JL, Walsh FB. Opsoclonus and ataxic conjugate movements of eyes. Arch Ophthalmol 1960;64:244-50.
- Vignaendra V, Lim CL. Electro-oculographic analysis of opsoclonus: Its relationship to saccadic and nonsaccadic eye movements. *Neurology* 1977;27:1129–33.
- 79. Glaser JS. Neuro-ophthalmology. Hagerstown MD: Harper & Row, 1978.
- Delreux V, Kevers L, Callewaert A, Sindic C. Opsoclonus secondary to an Epstein-Barr virus infection [letter]. Arch Neurol 1989;46:480-1.
- 81. Shetty T, Rosman NP. Opsoclonus and hydrocephalus. Arch Ophthalmol 1972;88:585-9.
- Tyczka W (cited by Malaguzzi-Valeri). O neizbomości mioklonicznej oczu (opsoklonja). Pol Gaz Lek 1925;4:967.
- Dropcho E, Payne R. Paraneoplastic opsoclonus-myoclonus. Association with medullary thyroid carcinoma and review of the literature. Arch Neurol 1986;43:410–15.
- Tal Y, Jaffe M, Sharf B, Amir N. Steroid-dependent state in a child with opsoclonus. J Pediatr 1983;103:420-1.
- Rivner MH, Jay WM, Green JB, Dyken PR. Opsoclonus in hemophilus influenzae meningitis. Neurology 1982;32:661-3.
- 86. Tamura K, Kuroiwa Y. Opsoclonus-myoclonus syndrome in acute cerebellar ataxia. Clinical and electrophyiological observations of an adult case. *Folia Psychiat Neurol Jap* 1971;25:129-35.
- 87. Dehaena I, Van Vleymen B. Opsoclonus induced by phenytoin and diazepam. Ann Neurol 1987;21:216.
- 88. Herman P. Opsoclonus and oscillopsia. Mt Sinai J Med (NY) 1981;48:412-7.
- 89. Gobbi G, Melideo G, Giovamardi R. The effect of intravenous L-5-HTP in the myoclonic encephalopathy of infants. *Neuropediatrics* 1986;17:63–5.
- 90. Eber AM, Krieger J, Monjour A, Collard M, Rohmer F. L'opsoclonus. Etude oculographique. [Opsoclonus. Oculographic study] Rev Neurol Paris 1977;133:559-70.
- Kilgo GR, Schwartze GM. Opsoclonus: Update on clinical and pathologic associations. J Clin Neuro-ophthalmol 1984;4:109–13.
- 92. Cogan DG. Opsoclonus. In: Vinken PJ, Bruyn GW, eds. Infections of the Nervous System, Part II. Amsterdam: North-Holland, 1978:611–17. Handbook of Clinical Neurology; Vol. 34.
- Gresty MA, Findley LJ, Wade P. Mechanism of rotatory eye movements in opsoclonus. Br J Ophthalmol 1980;64:923-25.
- 94. McLatchie GR, Young DD. Presenting features of thoracic neuroblastoma. Arch Dis Child 1980; 55:958.
- 95. Noda S, Takao A, Itoh H, Umezaki H. Opsoclonus in hyperosmolar nonketotic coma. J Neurol Neurosurg Psychiatry 1985;48:1186.

- 96. Marmion DE, Sandalands J. Opsoclonia: A rare sign in polioencephalitis. Lancet 1947;2:508-9.
- 97. Baker ME, Kirbs DR, Korobkia M, Bowie JD, Filston HC. The association of neuroblastoma and myoclonic encephalopathy: An imaging approach. *Pediatr Radiol* 1985;15:185–9.
- Latchaw RE, L'Heureux PR, Young G, Priest JR. Neuroblastoma presenting as central nervous system disease. Am J Neurol Res 1982;3:623-30.
- Ross AT, Zeman W. Opsoclonus, occult carcinoma, and chemical pathology in dentate nuclei. Arch Neurol 1967;17:546–51.
- 100. Kinast M, Levin HS, Rothner DA, Erenberg G, Wacksman J, Judge J. Cerebellar ataxia, opsoclonus and occult neural crest tumor. Am J Dis Child 1980;134:1057-59.
- 101. Blue SK, Janeway R, Stanley JA. Opsoclonus and body tremulousness. A case report with suggested cause. *Trans Am Neurol Assoc* 1971;96:208–10.
- Digre K. Opsoclonus in adults. Report of three cases and review of the literature. Arch Neurol 1986;43:1165-75.
- 103. Sandok BA, Kranz H. Opsoclonus as the initial manifestation of occult neuroblastoma. Arch Ophthalmol 1971;86:235-6.
- 104. Hashiba Y, Kuroda Y, Neshige R, Shibasaki H. A case of opsoclonus-polymyo-clonia syndromesuppression by TRH and 5-hydroxytryptophan. *Rinsho Shinkeigaku* 1984;24:1091–4.
- 105. Gastaut H. Semeiologie des myoclonies et nosologie analytique de syndromes myocloniques. [Semeiology of myoclonia and analytic nosology of myoclonic syndromes] Rev Neurol (Paris) 1968;119:1.
- Kalmanchey R, Veres E. Dancing eyes syndrome—brainstem acoustic evoked potential approach. Neuropediatrics 1988;19:193–6.
- 107. Gumbinas M, Gratz ES, Johnston GS, Schwartz AD. Positive gallium scan in the syndrome of opsoclonus-myoclonus treated with adrenocorticotropic hormone. *Cancer* 1984;54:815-6.
- Wilkins DE, Hallett M, Erba G. Primary generalized epileptic myoclonus: A frequent manifestation of central origin. J Neurol Neurosurg Psychiatry 1985;48:506-16.
- 109. Farrelly C, Daneman A, Chan HSL, Martin DJ. Occult neuroblastoma presenting with opsomyoclonus: Utility of computed tomography. AJR 1984;142:807-10.
- 110. Ziter FA, Bray PF, Cancilla PA. Neuropathological findings in a patient with neuroblastoma and myoclonic encephalopathy. Arch Neurol 1979;36:51.
- 111. Harada Y, Ishimitsu H, Nishimoto K, Miyata I, Matsumi N. Opsoclonus-polymyoclonia syndrome suppressed with reserpine. No To Shinkei 1986;38:359-62.
- 112. Mitra A, Bajaj S. Opsoclonus-myoclonus in acute cerebellar ataxia. J Assoc Physicians India 1987;35:853-5.
- 113. Sachdeva JR, Singh N, Mann GS. Acute cerebellar ataxia with opsoclonus. J Assoc Physicians India 1984;32:460.
- 114. Karobkin M, Clark RE, Palubinskas AJ. Occult neuroblastoma and acute cerebellar ataxia in childhood. *Radiology* 1972;102:151-2.
- 115. Kaplan M, Salet J, Pean G, Guran P, Straus P, Davy M. Sur une variété d'ataxie cérébelleuse acquise de l'enfance avec tremblement oculaire. [On a variety of cerebellar ataxia with eyeball tremors, acquired in childhood] *Arch Franc Pediatr* 1959;16:1124–29.
- 116. Weiss S, Carter S. Course and prognosis of acute cerebellar ataxia in children. *Neurology* 1959;9:711-21.
- Nickerson BG, Hutter JJ. Opsomyoclonus and neuroblastoma: Response to ACTH. Clin Pediatr 1979;18:446–8.
- 118. Winkler GF, Baringer JR, Sweeney VP, Cogan DG. An acute syndrome of ocular oscillations and truncal ataxia. *Trans Am Neurol Assoc* 1966;91:96–9.
- 119. Dailey AT, Pranzatelli MR. The association of opsoclonus, myoclonus, and ataxia: A review of the literature [Abstract]. Child Neurol Soc 1989;225.
- 120. Klingman WO, Hodges FG. Acute ataxia of unknown origin in childhood. J Pediatr 1944;24: 536-43.
- 121. Keller MJ, Karelitz S. Acute ataxia in a twenty-month-old female. Pediatrics 1948;1:754-57.
- 122. Blaw ME, Sheedan JC. Acute cerebellar syndrome of childhood. Neurology 1958;8:538-42.
- King G, Schwartz GA, Slade HW. Acute cerebellar ataxia of childhood. *Pediatrics* 1958;21:731– 45.
- 124. Lasater GM, Jabbour JT. Acute ataxia of childhood: A summary of fifteen cases. Am J Dis Child 1959;97:61-5.
- 125. Berg R, Jelke H. Acute cerebellar ataxia in children associated with Coxsackie viruses group B. Acta Paediatr Scand 1965;54:497–502.
- 126. Aikawa T, Takemiya T, Kobayoshi I, et al. An acute syndrome of opsoclonus and body tremulousness: A case of benign encephalitis. No To Shinkei 1984;36:121-6.

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- 127. Dalrymple JM. Opsoclonus and generalized tremulousness. Clini-Pearls 1980;3:6-7.
- 128. Sheinman BD, Gawler J. Opsoclonus and polymyoclonia complicating oat-cell carcinoma of the bronchus. *Postgrad Med J* 1982;58:704–5.
- 129. Graus F, Cordon-Cardo C, Cho E-S, Posner J. Opsoclonus and oat cell carcinoma of the lung: Lack of evidence for anti-CNS antibodies. *Lancet* 1984;1(8392):1479.
- 130. Berglund G, Mossberg HO, Rydenstam B. Acute cerebellar ataxia in children. Acta Paediatr Upps 1955;44:254-62.
- 131. Ellenberger C, Campa JF, Netsky MG. Opsoclonus and parenchymatous degeneration of the cerebellum: The cerebellar origin of an abnormal ocular movement. *Neurology* 1968;18:1041-6.
- 132. Lechi A, Tedeschi F, Terzano MG, Trabattoni G. Opsoclonus and myoclonus in malignant disease. A case report. Acta Neurol (Napoli) 1983;5:205-9.
- 133. Kearsley JH, Johnson P, Halmagyi GM. Paraneoplastic cerebellar disease. Remission with excision of the primary tumor. *Arch Neurol* 1985;42:1208–10.
- 134. Mitchell WG, Snodgrass SR. Opsoclonus-ataxia due to childhood neural crest tumors; a chronic neurologic syndrome. J Child Neurol 1990;5:153-8.
- 135. Malaguzzi-Valeri O. Su una rara sindrome clonica degli occhi e del muscoli scheletrici (oftalmomioclonia) in una caso di attasia cerebellare ocula. [On a rare clonal syndrome of the eyes and skeletal muscles (Ophthalmo Myoclonia) in a case of acute cerebellar ataxia] *Riv Patol Nerv Ment* 1939;54:103.
- 136. Sabouraud O, Chatel M, Badiche A, et al. Un cas d'encéphalite avec opsoclonies. [A case of encephalitis with opsoclonia] Rev Otoneuro-Ophthalmol 1973;45:139-44.
- 137. Forster C, Lenard HG, Pache HD, Versmold H. Die infantile myoklonische encephalopathie. [Infantile myoclonic encephalopathy] Z Kinderheilkd 1971;111;67–82.
- 138. Gross-Selbeck G, Goll U. Myoclonic encephalopathy ("Dancing eyes" syndrome with myoclonic ataxia). *Klin Pädiatr* 1972;184:393–9.
- 139. Heyer R, Hamann J, Offner G, Uebbing G. Die myoklonische Encephalopathie im Kindesalter. [Myoclonic encephalopathy in childhood] *Monatsschr Kinderheilkd* 1977;125:640–5.
- 140. Canedo JA, Hupp SL, Hamilton WJ. Opsoclonus and ataxia accompanying acute gastroenteritis. *Ala Med* 1988;57:39-44.
- 141. Vieira JBA, Rosa ED. Polymioclonia-opsoclono: Sindrome de Kinsbourne. Relato de um caso [palymyoclonia-opsoclonus: Kinsbourne's syndrome. A case report]. Arq Neuropsiquiatr 1985; 43:194–7.
- 142. Ruangkit C, Limapichat K. Opsoclonus in mumps and poliovirus type 3 encephalitides: A report of 2 cases. J Med Assoc Thailand 1989;72:417-20.
- 143. Ichiba N, Miyake Y, Sato K, Oda M, Kimoto H. Mumps-induced opsoclonus-myoclonus and ataxia. *Pediatr Neurol* 1988;4:224-7.
- 144. Brewis EG. Viral meningo-encephalitis. Ciba Foundation Study Group 1960; No. 7:46.
- 145. Strickland B. Poliomyelitis. Lancet 1947;2:369.
- 146. Arthius M, Lyon G, Thieffray ST. La forme ataxique de la maladie de heine-medin. [The ataxic form of heine-medin disease] *Rev Neurol* (*Paris*) 1960;103:329-40.
- 147. Curnen EC, Chamberlin HR. Acute cerebellar ataxia associated with poliovirus infection. Yale J Biol Med 1961;34:219-33.
- 148. Krebs E, Messimy R, Petit-Dutaillis D. Clonies bilaterales en salves des globes oculaires et des orteils associées à des troubles de l'équilibre, apparus lors d'une réchute d'un syndrome infectieux avec ictère. [Bilateral clonisms in salvos of the eyeballs and toes associated with balance disorders appearing during a recurrence of an infectious syndrome with jaundice] *Rev Neurol* (*Paris*) 1958:3-33.
- 149. Vejjajiva A, Lerdverasirikul P. Opsoclonus in salmonella infection. Br Med J 1977;2:1260.
- 150. Evans RW, Welch KWA. Opsoclonus in a confirmed case of St. Louis encephalitis. J Neurol Neurosurg Psychiatr 1982;45:660-1.
- 151. Southern PM Jr, Smith JW, Luby JP, Barnett JA, Sanford JP. Clinical and laboratory features of epidemic St. Louis encephalitis. Ann Intern Med 1969:681-9.
- 152. Kumar N, Singh S. Opsoclonus in tuberculous meningitis. J Assoc Physicians India 1989;37: 353-4.
- 153. Yagi S, Moriya O, Nakajima M, Umeki S, Hino J, Soejima R. A case of tuberculous pleurisy associated with myoclonus and Quincke's edema due to isoniazid and isoniazid sodium methanesulfonate. *Kekkaku* 1989;64:407–42.
- 154. Asindi AA, Bell EJ, Browning MJ, Stephenson JB. Vaccine-induced polioencephalomyelitis in Scotland. Scot Med J 1988;33:306–7.

- Kennedy C, Tucket SA. Ocular myoclonus: A clinical and pathological study. *Neurology* 1962; 12:297–280.
- 156. Thieffry St, Martin Ch, Arthuis M. Les formes encéphaliques de la maladie de heine-medin. Sem Hop Paris 1959;35:259.
- 157. Lucking CH, Birnberger KL, Straschill M. Opsoclonus, truncal ataxia and myoclonus as signs of a benign brainstem encephalitis. *Electroencephalogr Clin Neurophysiol* 1975;39:212.
- 158. Zangemeister VH, Muller-Jensen A, Zschocke S. Benign encephalitis: Electro-oculographic analysis of opsoclonus. J Neurol 1979;222:95-108.
- 159. McAllister RM. Neuroblastoma: A viral etiology? J Pediatr Surg 1968;3:138.
- 160. Dodge PR. In: Bray PF, Ziter F, Lahey ME, Myers G, eds. The coincidence of neuroblastoma and acute cerebellar encephalopathy. *Trans Am Neurol Assoc* 1969;94:106-9.
- 161. Arnason BGW. Paraneoplastic syndromes of muscle, nerve, and brain: Immunologic considerations. In: Rose FC, ed. Clinical neuroimmunology, London: Blackwell, 1979:421–50.
- 162. Dropcho EJ. The remote effects of cancer on the nervous system. Neurol Clin 1989;7:579-603.
- Brain WR, Wilkinson M. Subacute cerebellar degeneration associated with neoplasms. Brain 1965;88:465–78.
- 164. Lambert EH, Rooke ED. Myasthenic state and lung cancer. In: Brain WL, Norris FH, eds. *Remote effects of cancer on the nervous system.* New York: Grune & Stratton, 1965:67–80.
- 165. Denny-Brown D. Primary sensory neuropathy with muscular changes associated with carcinoma. J Neurol Neurosurg Psychiatry 1948;11:73-87.
- 166. Henson RA, Russel DS, Wilkinson M. Carcinomatous neuropathy and myopathy. A clinical and pathological study. Brain 1954;77:82-121.
- 167. Croft PB, Wilkson M. The incidence of carcinomatous neuromyopathy with special reference to carcinoma of the lung and the breast. In: Brain WL, Norris FH, eds. Remote effects of cancer on the nervous system, New York: Grune & Stratton, 1965:44-54.
- 168. Henson RA, Hoffman HL, Urich H. Encephalomyelitis with carcinoma. Brain 1965;88:449-64.
- Corsellis JAN, Goldberg GJ, Norton AR. Limbic encephalitis and its association with carcinoma. Brain 1981;91:481-95.
- 170. Albin RL, Bromberg MB, Penney JB, Knapp R. Chorea and dystonia: A remote effect of carcinoma. *Mov Disord* 1988;2:162–9.
- 171. Keane JR, Devereaux MW. Opsoclonus associated with an intracranial tumor and clinicopathologic case report. Arch Ophthalmol 1974;92:443-5.
- 172. Bellur SN. Opsoclonus: Its clinical value. Neurology 1975;25:502.
- 173. Keating JW, Cromwell LD. Remote effects of neuroblastoma. AJR 1978;131:299-303.
- 174. Alessi D. Lesioni parenchimatose del cervelletto da carcinoma uterino (gliosi carcinotossica?): Sintomatologia dissinergico-mioclonica. [Parenchymatous lesions of the cerebellum from uterine carcinoma (carcinotoxic neuroglioses): Dyssynergic-myoclonic symptomatology] Riv Pat Nerv Ment 1940;55:148-174.
- 175. Cushing H, Wolbach SB. The transformation of a malignant paravertebral sympathicoblastoma into a benign ganglioneuroma. Am J Pathol 1927;3:302.
- Larbrisseau A, Geoffroy G, Lasalle R. Association du'un neuroblastome avec un syndrome de polymyoclonie-opsoclonie. Un Med Can 1973;102:1691-94.
- 177. Pochedly C, Strauss L. Histogenesis and pathology of Neuroblastoma. In: Pochedly C, ed. Neuroblastoma, Acton MA: Publishing Sciences, 1976:155-79.
- 178. Young DG. Thoracic neuroblastoma/ganglioneuroma. J Pediatr Surg 1983;18:37.
- 179. Green AA, Hayes FA. Neuroblastoma. In: Rudolph AM, Hoffman JIE, Axelrod S, eds. Pediatrics, Norwalk CT: Appleton and Lange 1987:1113-6.
- 180. Clatworthy HW. The treatment of neuroblastoma. CA 1968;3:146-150.
- 181. Dargeon HW. Neuroblastoma. J Pediatr 1962;61:456.
- 182. Hohischneider AM, Geiger H, Bolkenius N, Janka G, Lampert F. Spätfolgen beim Neuroblastom: Paraneoplastische Erkrankungen und Therapiefolgen [Neuroblastoma: Paraneoplastic disease and late complications] *Monatsschr Kinderheilkd* 1977;125:69–73.
- 183. Stowens D. Tumors of the peripheral nervous system and neural crest derivatives. In: *Pediatric Pathology*, 2nd Ed, Baltimore: Williams & Wilkins, 1966:412-23.
- 184. Bove K, McAdams J. Composite galgioneuroblastoma. Arch Pathol Lab Med 1981;105:325-30.
- 185. Adam A, Hochholzer L. Ganglioneuroblastoma of the posterior mediastinum. Cancer 1981;47: 373-81.
- 186. Donohue JP, Garrett RA, Baehner RL. The multiple manifestations of neuroblastoma. Trans Am Assoc Genitourin Surg 1973;65:102-6.
- 187. Beckwith JB, Perrin EV. In situ neuroblastomas: A contribution to the natural history of neural crest tumors. Am J Pathol 1963;43:1089-1104.

- 188. Everson TC, Cole WH. Spontaneous regression of cancer. Philadelphia: Saunders, 1966:88.
- 189. Bill AH. The regression of neuroblastoma. J Pediatr Surg 1968;3:103.
- Ziegelbaum MM, Kay R, Rothner AD, Lorig R. The association of neuroblastoma with myoclonic encephalopathy of infants: The use of magnetic resonance as an imaging modality. J Urol 1988;139:81-2.
- 191. Berger PE, Kukn JP, Munschauer RW. Computed tomography and ultrasound in the diagnosis and management of neuroblastoma. *Radiology* 1978;128:663-7.
- 192. Donaldson JS, Gilsanz V, Miller JH. CT scanning in patients with opso-myoclonus: Importance of nonenhanced scan. AJR 1986;146:781-3.
- 193. Armstrong EA, Harwood-Nash DCF, Fitz CR, Chuang SH, Pettersson H, Martin DJ. CT of neuroblastomas and ganglioneuromas in children. AJR 1981;139:571-6.
- 194. Siegel MJ, Sagei SS. Computed tomography as a supplement to urography in evaluation of suspected neuroblastoma. *Radiology* 1982;142:435–8.
- 195. Roberts KB, Freeman JM. Cerebellar ataxia and "occult neuroblastoma" without opsomyoclonus. *Pediatrics* 1975;56:464-5.
- Bekerman C, Port RB, Pang E, Moohr JW, Kranzler JK. Scintigraphic evaluation of childhood malignancies by ⁶⁷Ga-citrate. *Radiology* 1978;127:719–25.
- 197. Kincaid OW, Hodgson JR, Dockerty MD. Neuroblastoma: A roentgenologic and pathologic study. Am J Roentgenol Radium Ther Nucl Med 1957;78:420.
- 198. Belasco JB, McMillan CW. Occult neuroblastoma: Difficulties of diagnosis. Am J Dis Child 1979;133:858.
- 199. Rupprecht L, Mortier W. Cerebellare Bewegungsstörung and Neuroblastoma. [Cerebellar motor disturbance and neuroblastoma] *Monatschr Kinderheilkd* 1975;123:392–3.
- Dropcho EJ, Obata Y, Old LJ, Posner JB. Isolation of a cDNA clone coding for a Purkinje cell protein related to paraneoplastic cerebellar degeneration. *Neurology* 1986;36(Suppl 1):331.
- 201. Maccario M, Seelinger D, Snyder R. Thallotoxicosis with coma and abnormal eye movements (opsoclonus): Clinical and EEG correlations. *Electroencephalogr Clin Neurophysiol* 1975;38:98.
- 202. Coldman AJ, Fryer CJH, Elwood JM, Sonley MJ. Neuroblastoma: Influence of age at diagnosis, stage, site, and sex on prognosis. *Cancer* 1980;46:1896–1901.
- 203. Filler RM, Traggis DG, Jaffe N, Vawter GF. Favorable outlook for children with mediastinal neuroblastoma. J Pediatr Surg 1972;7:136-43.
- 204. Jereb B, Bretsky SS, Vogel R, Helson L. Age and prognosis in neuroblastoma. Review of 112 patients younger than 2 years. Am J Pediatr Hematol Oncol 1984;6:233-43.
- Bachmann K-D. Das Neuroblastoma sympathicum. Klinische prognose von 1030 fallen. [Sympathiconeuroblastoma. Clinical (course and) prognosis of 1030 cases] Z Kinderheilkd 1962;86: 710-24.
- Evans AE, D'Angio GJ, Randolph J. A proposed staging for children with neurblastoma. Cancer 1971;27:374–8.
- 207. Brodeur GM, Seeger RC, Barrett A, et al. International criteria for diagnosis, staging and response to treatment in patients with neuroblastoma. In: Evans AE, D'Angio GJ, Knudson AG, Seeger RC, eds. Advances in neuroblastoma research 2. New York: Alan R Liss, 1988:509-24.
- 208. Lipinski Ch, Kratzer W, Daum R. Die infantile myoklonische Enzephalopathie—ein paraneoplastisches Syndrom bei Neuroblastom. [Myoclonic encephalopathy of infants—a paraneoplastic syndrome in neuroblastoma] Z Kinderchir 1975;16:111–17.
- 209. Cottom DG. Acute cerebellar ataxia. Arch Dis Child 1957;32:181-8.
- Donohue JP, Garrett RA, Baehner RL, Thomas MH. The multiple manifestations of neuroblastoma. J Urol 1974;111:260–4.
- 211. Au WJ, Keltner JL. Opsoclonus with amitriptyline overdose. Ann Neurol 1979;6:87.
- Morres CA, Dire DJ. Movement disorders as a manifestation of nonketotic hyperglycemia. J Emerg Med 1989;7:359-64,1989.
- 213. Burks JS, Walker JE, Rumack BH, et al. Tricyclic antidepressant poisoning: reversal of coma, choreoathetosis, and myoclonus by physostigmine. JAMA 1974;230:1405-7.
- 214. Noble J, Matthew H. Acute poisoning by tricyclic antidepressants: Clinical features and management of 100 patients. *Clin Toxicol* 1969;2:403-21.
- 215. Taylor JR, Selhoist JB, Houff SA, Martinez AJ. Chlordecone intoxication in man. *Neurology* 1978;28:626-30.
- 216. Scharf D. Opsoclonus-myoclonus following the intranasal usage of cocaine. J Neurol Neurosurg Psychiatry 1989;52:1447-8.
- 217. Garcin MR, Ginsbourg M, Godlewski ST, Emile J. Intoxication par le DDT: Syndrome meningoencéphalique aigu regressif avec decharges cloniques diffuses et mouvements désordonnés "en salves" des globes oculaires. [Intoxication with DDT: Acute regressive meningoencephalitic

syndrome with diffuse clonal discharges and disordered "salvo" movements of the eyeballs] Rev Neurol (Paris) 1965;113:559-64.

- 218. Cohen WJ, Cohen NH. Lithium carbonate, haloperidol, and irreversible brain damage. JAMA 1974;230:1283-7.
- 219. Favarel-Garriques B. Two cases of severe poisoning by lithium carbonate. Am Med Psychol 1972;1:253.
- 220. Rosen PB, Stevens R. Action myoclonus in lithium toxicity. Ann Neurol 1983;13:221-2
- 221. Pullicino P, Aquilina J. Opsoclonus in organophosphate poisoning. Arch Neurol 1989;46:704-5. 222. Lazar RB, Ho SU, Melen O, Daghestani AN. Multifocal central nervous system damage by
 - toluene abuse. *Neurology* 1983;33:1337.
- 223. Supin-Viterbo V, Sicard C, Risvegliato M, Rancurel G, Buge A. Toxic encephalopathy due to ingestion of bismuth salts: Clinical and EEG studies of 45 patients. J Neurol Neurosurg Psychiatry 1977;40:748-52.
- 224. Molina JA, Calandre L, Bermejo F, Posadas F, Fernandez-Ortega yJD. Encefalopatia mioclonica por sales de bismuto. Eficacia del tratamiento con dimercaprol. [Myoclonic encephalopathy caused by bismuth salts. Effectiveness of dimercaprol therapy]. *Med Clin (Barc)* 1989;93:20–2.
- Rouzaud M, Degiovanni E, Delplace MP, Lemaire JF, Paulin M. L'encéphalopathie avec opsoclonies. [Encephalopathy with opsoclonia (on an observation)] Oto-Neuro-Ophthalmol 1976;48: 43-55.
- 226. Lucas C, Soetaert G, Leys D, Petit H. Myoclonus during a treatment with buflomedil. Acta Clin Belg 1989;44:360-1.
- 227. Gottwald W Van. Transitoriches Abheilen von Psoriasis-Effloreszenzen wahrend Hydantoinvergiftung mit seltener zentralnervöser Symptomatik. [Transitory healing of psoriasis efflorescence during hydantoin intoxication with rare central nervous system symptoms] *Derm Wochenschr* 1968;154:241.
- 228. Wendland KL. Myoklonien nach gaben von carbamazepin. Nervenarzt 1969;389:231.
- 229. Moene PMMY, Cuche M, Trillet M, Motin J, Michael D. Problèmes diagnostiques poses par l'intoxication aigue au chloralose (à propos de 6 cas) [Diagnostic problems set by acute chloralose intoxication in connection with six cases]. J Med Lyon 1969;50:1483.
- 230. Ammenti A, Reitter B, Muller-Wiefel DE. Chlorambucil neurotoxicity: Report of two cases. *Helv Paediatr Acta* 1980;35:281–7.
- 231. Ng ATH. Prolonged myoclonic contractions after enflurane anesthesia—a case report. Can Anaesth Soc J 1980;27:502–3.
- 232. Helmers JH, Adam AA, Giezen J. Pain and myoclonia during induction with etomidate. Acta Anaesth Belg 1981;32:141-7.
- Pfeiffer CC, Jenney EH, Marshall WH. Experimental seizures in man and animals with acute pyridoxine deficiency produced by hydrazides. *Electroencephalogr Clin Neurophysiol* 1956;8: 307.
- 234. Westheimer R, Klawans HL. The role of serotonin in the pathophysiology of myoclonic seizures associated with acute imipramine toxicity. *Neurology* 1974;24:1175–7.
- 235. Klawans HL, Goetz C, Bergen D. Levodopa-induced myoclonus. Arch Neurol 1975;32:331-4.
- 236. Vardi J, Glaubman H, Rabey JM, Streifler M. Myoclonic attacks induced by L-dopa and bromocriptine in Parkinson patients. J Neurol 1978;218:35-42.
- 237. Aigner BR, Mulder DW. Myoclonus. Clinical significance and an approach to classification. Arch Neurol 1960;2:600.
- 238. Mack RB. Toxic encounters of the dangerous kind: Methaqualone intoxication. N Carol Med J 1981;42:796.
- Langlois M, Vercel RM, Kahlil, Bereni R. Sur quatre cas d'intoxication collective par le bromure de methyle. Considerations cliniques et électroéncephalographiques. [On four cases of collective intoxication with methylbromide. Clinical and electroencephalographic considerations] Rev Neurol (Paris) 1963;108:305.
- 240. Hyser CL, Drake ME. Myoclonus induced by metoclopramide therapy. Arch Intern Med 1983; 143:2201-02.
- 241. Potter JM, Reid DB, Shaw RJ, Hackett P, Hickman PE. Myoclonus associated with treatment with high doses of morphine: The role of supplemental drugs. *Br Med J* 1989;299:150–3.
- 242. Reutens DC, Stewart-Wynne EG. Norpethidine induced myoclonus in a patient with renal failure [letter]. J Neurol Neurosurg Psychiatry 1989;52:1450–1.
- 243. Kurtzman NA, Rogers P, Harter HR. Neurotoxic reaction to penicillin and carbenicillin. JAMA 1970;214:1320.
- 244. Hardin JA, Griggs RC. Diazepam treatment in a case of strychnine poisoning. Lancet 1971;2:372.

- 245. Lane JC, Tennison MG, Lawless ST, Greenwood RS, Zaritsky AL. Movement disorder after withdrawal of fentanyl infusion. J Pediatr 1991;119:649-51.
- 246. Burns RS, Lerner SE, Corrado PA, James SH, Schnoll SH. Phencyclidine-states of acute intoxication and fatalities. West J Med 1975;123:345-9.
- Chaptal J, Jean R, Labauge R, Bonnet H, Aghai E. Myoclonies oppositionnelles par intoxication à la piperazine. [Oppositional myoclonia due to piperazine intoxication] Arch Fr Pediatr 1963; 20:17.
- 248. Cohen RM, Pickar D, Murphy DL. Myoclonus-associated hypomania during MAO-inhibitor treatment. Am J Psychiatr 1980;137:105-6.
- De Grood PM, Harbers JB, Van Egmond J, Crul JF. Anaesthesia for laparoscopy. A comparison of five techniques including propofol, etomidate, thiopentone and isoflurane. *Anaesthesia* 1987; 42:815-23.
- 250. Castillo-Monsequr J, Villalonga-Morales A, Nalda-Felipe MA. [Pharmacologic prevention of myoclonia during anesthesia induction with etomidate. Comparative study of fentanyl, fluni-trazepam, and pancuronium.] *Rev Esp Anetesiol Reanim* 1987;34:270–2.
- 251. Aguglia U, Zappia M, Quattrone A. Carbamezepine-induced nonepileptic myoclonus in a child with benign epilepsy. *Epilepsia* 1987;28:515-8.
- 252. White PD. Myoclonus and episodic delirium associated with phenelzine: A case report. J Clin Psychiatry 1987;48:340-1.
- 253. Casas M, Garcia-Ribera C, Alvarez E, Udina C, Queralto JM. Myoclonic movements as a side effect of treatment with therapeutic doses of clomipramine. *Int Clin Psychopharmacol* 1987;2: 333–6.
- 254. Edwards DL, Johnson CE. Insect-repellant-induced toxic encephalopathy in a child. *Clin Pharm* 1987;6:496–8.
- Bowdle TA. Myoclonus following suferianil without EEG seizure activity. Anesthesiology 1987; 67:593-5.
- 256. Barsan WG, Olinger CP, Adams HP, et al. Use of high dose naloxone in acute stroke: Possible side-effects. Crit Care Med 1989;17:762-7.
- 257. Uchihara T, Tsukagoshi H. Myoclonic activity associated with cefmetazole, with a review of neurotoxicity of cephalosporins. Clin Neurol Neurosurg 1988;90:369-71.
- 258. Little JT, Jankovic J. Tardive myoclonus. Mov Disord 1987;2:307-11.
- 259. Eisele G. Neurologic complications of metoclopramide therapy [letter]. NY State J Med 1988; 88:332.
- Madigand M, Dien J, Pavin G, Allain H. [Myoclonic encephalopathy probably attributable to propafenones (letter)]. Presse-Med 1988;17:538.
- 261. Lippman S, Moskovitz R, O'Tuama L. Tricyclic-induced myoclonus. Am J Psychol 1977;134: 90-1.
- Parker WD, Goodman SI, Stumpf DA. Biotin responsive opsoclonus-myoclonus syndrome. Neurology 1983;33(Suppl 2):153.
- Hoyt CS, Mousel DK, Weber AA. Transient supranuclear disturbances of gaze in healthy neonates. Am J Ophthalmol 1980;89:708-13.
- 264. Bienfang DC. Opsoclonus in infancy. Arch Ophthalmol 1974;91:203-5.
- 265. Brichet B, Andre JM, Reny A, Weber M. Etude électromyographique d'opsoclonies survennes au cours d'un syndrome d'insuffisance vertébrobasilaire. [Electromyographic study of opsoclonia occurring during a basilar artery insufficiency syndrome] Rev Neurol (Paris) 1970;122:439– 43.
- 266. Arnould G, Laxenaire M, Picard L, Andre JM, Brichet B. Les encephalopathies avec opsoclonies. Révue générale à propos d'un cas par insuffisance vertébro-basilaire. Ann Med Nancy 1970;9:587–95.
- Chowdhary UM, Ibrahim AW, Ammar AS, Dawodu AH. Tecto-cerebellar dysraphia with occipital encephalocele. Surg Neurol 1989;31:310–14.
- Mattyus A, Veres E. Multiple sclerosis in childhood: Long-term katamnestic investigations. Acta Pediatr Hung 1985;26:193-204.
- 269. Coppeto JR, Monteiro ML. Craniofacial dysmorphism and opsoclonus. Ophthalmic Paediatr Genet 1984;4:147-153.
- 270. Fierro M, Martinez AJ, Harbison JW, Hay SH. Smith-Optiz syndrome: Neuropathological and ophthalmological observations. *Dev Med Child Neurol* 1977;19:57-62.
- 271. Joubert M, Eisenring JJ, Robb JP, Andermann F. Familial agenesis of the cerebellar vermis. A syndrome of episodic hyperpnea, abnormal eye movements, ataxia and retardation. *Neurology* 1969;19:813-25.
- 272. Keane JK. Transient opsoclonus with thalamic hemorrhage. Arch Neurol 1980;37:423-4.

- Weissman B, Devereaux M, Chandar K. Opsoclonus and hyperosmolar stupor. *Neurology* 1989; 39:1401–2.
- 274. Schou HI. Myoklonus-mit eigentumlichen Gehirnveränderungen. Z Ges Neurol Psychol 1925; 95:12-20.
- 275. Turazzi S, Alexander A, Bricolo A, Rizzuto N. Opsoclonus and palatal myoclonus during prolonged post-traumatic coma. A clinico-pathologic study. *Eur Neurol* 1977;15:257.
- 276. Robin A, Vallat M, Tapie P, Vallat J-N. Opsoclonus et troubles neurologiques associés [Opsoclonus and associated neurological disorders]. Arch Ophthalmol (Paris) 1976;36:645-56.
- 277. Tahmoush AJ, Brooks JE, Keltner JL. Palatal myoclonus associated with abnormal ocular and extremity movements. *Arch Neurol* 1927;27:431.
- 278. Matsuo F, Ajax ET. Palatal myoclonus and denervation supersensitivity in the central nervous system. Ann Neurol 1972;5:72.
- Kalashnikova LA, Lavrova SV. [Soft palate myoclonus and opsoclonus in disorders of cerebrovascular circulation in the vertebrobasilar system.] *Zh Nevropatol Psikhiatr* 1989;89:100–5.
- 280. Guillain G. The syndrome of synchronous and rhythmic palato-pharyngo-laryngo-oculodiaphragmatic myoclonus. Proc R Soc Med 1938;31:1031-8.
- 281. Myers GJ. The therapy of myoclonus. In: Charlton MH, ed. *Myoclonic seizures*. Princeton: Excerpta Medica, 1975:121-60.
- 282. Carlow TJ. Medical treatment of nystagmus and ocular motor disorders. Int Ophthalmol Clin 1986;26:251-64.
- 283. Bale JF, Bray PF. Convulsions with an occult neural-crest tumor [letter]. N Engl J Med 1979; 301:555.
- 284. Telander RL, Smithson WA, Groover RV. Clinical outcome in children with acute cerebellar encephalopathy and neuroblastoma. J Pediatr Surg 1989;24:11-4.
- 285. Nichelli P, Bahmanian-Behbahani G, De Pellegrino G. [Opsoclonus-ataxia syndrome. Description of a case] *Riv Patol Nerv Ment* 1984;105:239-48.
- 286. Jabbari B, Urban E. Abnormal visual-evoked responses and opsoclonus. J Clin Neuro-Ophthalmol 1981;1:269-71.
- 287. Wolpow ER, Richardson ED. Case records of the Massachusetts General Hospital. Case 9-1988. A 57-year-old woman with worsening opsoclonus. N Engl J Med 1988;318:563-70.
- 288. Tuchman RF, Alvarez LA, Kantrowitz AB, Moser FG, Llena J, Moshe SL. Opsoclonusmyoclonus syndrome: Correlation of radiographic and pathological observations. *Neuroradiol*ogy 1989;31:250-52.
- Bostrom B, Mirkin BL. Elevation of lumbar cerebrospinal fluid catecholamine metabolites in patients with cranial and/or intracranial metastic neuroblastoma. In: Evans AE, D'Angio GJD, Knudson AG, Seeger RC, eds. Advances in neuroblastoma research 2, New York: Alan R Liss, 1988:557-64.
- 290. Pranzatelli MR, Huang Y-Y, Stanley M, Tate ED, Noetzel MJ, Lange BM. Cerebrospinal fluid monoamines in opsoclonus-myoclonus [Abstract]. Ann Neurol 1991;30:503.
- 291. Lema F, Maison A, Roseto A, Driencourt M, Piussan Ch. Présence d'interféron dans le LCR au cours d'une ataxie opsomyoclonique d'origine nontumorale. [Presence of interferon in the CSF of a child with nontumoral cerebellar taxia] Arch Fr Pediatr 1983;40:331–3.
- 292. Ninane J, Vermylen C, Cornu G. Tumour markers in neuroblastoma. In: Sluyser M, Voute A, eds. *Molecular biology and genetics of childhood cancers: Approaches to neuroblastoma*, Chichester: Ellis Horwood, 1988:13-25.
- 293. Hoefnagel CA, Voute PA, de Kraker J, Marcuse HR. Radionuclide diagnosis and therapy of neural crest tumors using iodine-131 meta-iodobenzylguanidine. J Nucl Med 1987;28:308-14.
- 294. Bergh J, Esscher T, Steinholtz L, et al. Immunocytochemical demonstration of neuron-specific enolase (NSE) in human lung cancers. Am J Clin Pathol 1985;84:1–7.
- 295. Mirkin B, Fink DW. Modulation of neuroblastoma growth by biological factors and pharmacological agents. In: Sluyser M, Voute PA, eds. *Molecular biology and genetics of childhood cancers: Approaches to neuroblastoma*, Chichester: Ellis Horwood, 1988:72–95.
- 296. Pegelow C, Ebbin AJ, Powars D, Towner J. Familial neuroblastoma. J Pediatr 1975;87:763-5.
- 297. Hecht F, Hecht BK, Northrup JC, Trachtenberg N, Wood TS, Cohen JD. Genetics of familial neuroblastoma: Long range studies. *Cancer Genet Cytogenet* 1982;7:227-30.
- 298. Nutt JG, Bird TD. Essential myoclonus in a kindred with familial malignant melanoma. Arch Neurol 1984;41:189–91.
- 299. Knudson AG, Strong LC. Mutation and cancer: Neuroblastoma and pheochromocytoma. Am J Hum Genet 1972;24:514-32.
- 300. Schwab M. Molecular genetics of human neuroblastoma. In: Sluser M, Voute A, eds. Molecular

biology and genetics of childhood cancers: Approaches to Neuroblastoma. Chichester: Ellis Horwood, 1988:25–37.

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- Kaneko Y, Kanda N, Maseko N, et al. Different karyotypic patterns in early and advanced stage neuroblastoma. Cancer Res 1987;47:311–8.
- 302. Warrier RP, Kini R, Besser A, Wiatrak B, Raju U. Opsoclonus and neuroblastoma. *Clin Pediatr* 1985;24:32-4.
- 303. Miller RW, Fraumeni JF, Hill JA. Neuroblastoma: Epidemiologic approach to its origin. Am J Dis Child 1968;115:253-61.
- Cohn SL, Salwen H, Herst CV, et al. Single copies of the N-myc oncogene in neuroblastomas from children presenting with the syndrome of opsoclonus-myoclonus. *Cancer* 1988;62:723–6.
- 305. Schweisguth O. Excretion of catecholamine metabolites in urine of neuroblastoma patients. J Pediatr Surg 1968;3:118-20.
- Gitrow SE, Bertani-Dziedzic L, Dziedzic S. Patterns of catecholamine metabolism in neuroblastoma. In: Pochedly C, ed. Neuroblastoma: Clinical and biological manifestations, New York: Elsevier, 1982:97-130.
- Kaser H. Catecholamine producing tumors other than pheochromocytoma. *Pharmacol Rev* 1966; 18:659.
- 308. LaBrosse EH, Com-Nougue C, Zucker JM, et al. Urinary excretion of 3-methoxy-4hydroxymandelic acid and 3-methoxy-4-hydroxyphenylacetic acid by 288 patients with neuroblastoma and related neural crest tumors. *Cancer Res* 1980;40:1995.
- 309. Geiser CF, Efron ML. Cystathioninuria in patients with neuroblastoma or ganglioneuroblastoma: Its correlation to manilmandelic acid excretion and its value in diagnosis and therapy. *Cancer* 1968;22:856-60.
- 310. Hecht ST, Brasch RC, Styne DM. CT localization of occult secretory tumours in children. *Pediatr Radiol* 1982;12:67-71.
- Harel S, Yurgenson U, Rechavi G, Burstein Y, Spirer Z. Cerebellar ataxia and opsoclonus as the initial manifestations of myoclonus encephalopathy associated with neuroblastoma. *Childs Nerv* Syst 1987;3:245-7.
- 312. Graham-Pole J, Salmi T, Anton AH, Abramowsky C, Gross S. Tumor and urine catecholamines (CATS) in neurogenic tumors. *Cancer* 1983;51:834–9.
- 313. Yoffe JR, Brochardt RT. Characterization of serotonin uptake in cultured neuroblastoma cells: Differences between differentiated and nondifferentiated cells. *Mol Pharmacol* 1982;21:362–7.
- 314. Bill AH, Morgan A. Evidence for immune reactions to neuroblastoma and future possibilities for investigation. J Pediatr Surg 1970;5:111-6.
- 315. Martin EF, Beckwith JB. Lymphoid infiltrates in neuroblastomas. Their occurrence and prognostic significance. J Pediatr Surg 1978;3:161.
- 316. Hellstrom IE, Hellstrom KE, Pierce GE, Bill AH. Demonstration of a cell bound and humoral immunity against neuroblastoma cells. *Proc Natl Acad Sci USA* 1968;60:1231.
- 317. Hellstrom IE, Hellstrom KE, Evans CA, Heppner GH, Pierce GE, Yang JPS. Serum-mediated protection of neoplastic cells from inhibition by lymphocytes immune to their tumor-specific antigens. *Proc Natl Acad Sci USA* 1969;62:362.
- 318. Kaliss N. The elements of immunologic enhancement: A consideration of mechanisms. Ann NY Acad Sci 1962;101:64.
- 319. Hellstrom I, Hellstrom KE, Bill AH, Pierce GE, Yang JPS. Studies on cellular immunity to human neuroblastoma cells. *Int J Cancer* 1970;6:172–88.
- 320. Robinson MJ, Howard RN. Neuroblastoma, presenting as myasthenia gravis in a child aged 3 years. *Pediatrics* 1969;43:111-3.
- 321. Cawley LP, James VL, Minarad BJ, Bryant SA. Antibodies to Purkinje cells and peripheral nerve in opsoclonia. *Lancet* 1984;10:509–10.
- 322. Stefansson K, Marton LS, Dieperink ME, Molnar GK, Schlaepfer WW, Helgason CM. Circulating autoantibodies to the 20,000 Dalton proteins of neurofilaments in the serum of healthy individuals. *Science* 1985;228:1117–9.
- 323. Plioplys AV, Greaves A, Yoshida W. Anti-CNS antibodies in childhood neurologic diseases. *Neuropediatrics* 1989;20:93–102.
- 324. Royal W, Galasko DK, McKhann GM, Cunningham JM, Dropcho EJ. Clinical course, immunological and biochemical features of a patient with paraneoplastic cerebellar dysfunction. *Neurology*
- 325. Greenlee JE, Brashear HR. Antibodies to cerebellar Purkinje cells in patients with paraneoplastic cerebellar degeneration and ovarian carcinoma. *Ann Neurol* 1983;14:609–13.
- 326. Jaeckle KA, Graus F, Houghton A, Cardon-Cardo C, Nielsen SL, Posner JB. Autoimmune

response of patients with paraneoplastic cerebellar degeneration to Purkinje cell cytoplasmic protein antigen. Ann Neurol 1985;18:592-600.

- 327. Budde-Steffen C, Anderson NE, Rosenblum MK, et al. An antineuronal autoantibody in paraneoplastic opsoclonus. Ann Neurol 1988;23:528-31.
- 328. Brown RH, Ronthal M, Cane S. Antibodies to 85,000 dalton protein in paracarcinomatous cerebellar degeneration. *Neurology* 1985;35:288.
- 329. Tanaka K, Yamazaki M, Sato S, Toyoshima I, Yamamoto A, Miyatake T. Antibodies to brain proteins in paraneoplastic cerebellar degeneration. *Neurology* 1986;36:1169–72.
- Greenlee JE, Brashear HR, Rodnitzky RL, Corbett JJ, Digre KB. Fall in antineuronal antibody titers and improvement of neurologic deficit following tumor removal in paraneoplastic cerebellar degeneration. *Neurology* 1986;36(Suppl 1):334.
- 331. Graus F, Elkon KB, Cordon-Cardo C, Posner JB. Sensory neuronopathy and small cell lung cancer. Antineuronal antibody that also reacts with the tumor. Am J Med 1986;80:45-52.
- 332. Graus F, Elkon KB, Lloberes P, et al. Neuronal antinuclear antibody (anti-Hu) in paraneoplastic encephalomyelitis simulating acute polyneuritis. Acta Neurol Scand 1987;75:249-52.
- 333. Grunwald GB, Klein R, Simmonds MA, Kornguth SE. Autoimmune basis for visual paraneoplastic syndrome in patients with small-cell lung carcinoma. *Lancet* 1985;1:658-61.
- 334. Luque FA, Furneaux HM, Ferziger R, et al. Anti-Ri: An antibody associated with paraneoplastic opsoclonus and breast cancer. Ann Neurol 1991;29:241-51.
- 335. Franz CN, Duerst RE, Ryan DH, Gelsomino NL, Constine LS, Gregory PK. Antineuroblastoma monoclonal antibodies which do not bind to bone marrow cells. In: Evans AE, D'Angio G, Seeger RC, eds. Advances in Neuroblastoma Research, 1985:485-99.
- 336. Danon YL, Garty BZ. Autoantibodies to neuroblastoma cell surface antigens in neuropsychiatric lupus. *Neuropediatrics* 1986;17:23-7.
- Trojanowski JQ, Lee VM-Y. Anti-neurofilament monoclonal antibodies: Reagents for the evaluation of human neoplasms. Acta Neuropathol 1983:155–8.
- 338. Garson JA, Beverly PCL, Coakham HB, Harper EI. Monoclonal antibodies against human T lymphocytes label Purkinje neurones of many species. *Nature* 1982;298:375–7.
- Herishanu Y, Apte R, Kuperman O. Immunologic abnormalities in opsoclonus cerebellopathy. Neuro-Ophthalmology 1985;5;271-5.
- 340. Pranzatelli MR, Kao PC, Tate ED, et al. Autoantibodies to ACTH in opsoclonus-myoclonus [Abstract]. Ann Neurol 1991;30:471-2.
- 341. Corrias A, Nurchi AM, Rossi G, Sorcinelli R, Pusceddu G, Corda R. Opsoclonic encephalopathy in childhood (Kinsbourne syndrome). *Pediatr Med Chir* 1985;7:437-41.
- 342. Landbeck G, Blaker G, Bock P, Kurme A, Wriedt K. Die zytostatische Behandlung maligner Tumoren in Kindesalter. Z Kinder-Chir 1969;6(Suppl):30.
- 343. Leigh RJ, Zee D. The Neurology of eye movements. Philadelphia: Davis, 1983.
- 344. Rohr W, Hohnstadt P, Janzen RWC, Muller-Jensen A, St. Zschnocke. Myoklonien als problem in der intensivmedizin. In: Mertens HG, Pruntek H, eds. *Pathologische* Erregbarkeit des Nervensystems und ihre Behandlung (verhandlungen der Deutschen Gesellschaft für Neurologie Band 1). New York: Springer, 1980:433–8.
- 345. Holland JT. Opsoclonus with myoclonus. Proc Aust Assoc Neurol 1975;12:161-5.
- 346. Obeso JA, Artieda J, Quinn N, et al. Piracetam in the treatment of different types of myoclonus. Clin Neuropharmacol 1988;11:529-36.
- 347. Pranzatelli MR, Snodgrass SR. The pharmacology of myoclonus. *Clin Neuropharmacol* 1985;8: 99–130.
- 348. Pranzatelli MR. The proposed role of neurotransmitter receptors in the pathophysiology of human myoclonic disorders. *Med Hypoth* 1989;30:55-60.
- 349. Witter A. On the presence of receptors for ACTH neuropeptides in the brain. In: Pepeu G, Juhar MJ, Enna SJ, eds. *Receptors for neurotransmitters and peptide hormones*. New York: Raven Press, 1980:407-14.
- 350. Pranzatelli MR. Effect of antiepileptic and antimyoclonic drugs on serotonin receptors in vitro. Epilepsia 1988;29:412-9.
- Trifiletti RR, Pranzatelli MR. Influence of ACTH and ACTH fragments on [³H]MK-801 binding to rat hippocampal NMDA receptors [Abstract]. Soc Neurosci 1990;16:785.
- 352. Terenius L. Effect of peptides and amino acids on dihydromorphine binding to the opiate receptor. J Pharm Pharmacol 1975;27:450-2.
- 353. Czlonkowski A, Hollt V, Herz A. Binding of opiates and endogenous opioid peptides to neuroleptic sites in the corpus striatum. *Life Sci* 1978;22:953-62.
- 354. Stengaard-Pedersen K, Larson LI. Interaction of putative opioid peptides with opiate receptors. Acta Pharmacol Toxicol 1981;48:39-46.

- 355. Snell CR, Snell PH. A molecular basis for the interaction of corticotrophin with opiate receptors. *FEBS Lett* 1982;137:209–12.
- 356. Braestrup C, Squires RF. Pharmacological characteristics of benzodiazepine receptors in the brain. Eur J Pharmacol 1978;48:263-70.
- 357. Schwyzer R. Organization and transduction of peptide information. *Trends Pharmacol Sci* 1980; 3:311-27.
- 358. Jolles J, Zwiers H, van Dongen CJ, Schotman P, Wirtz KWA, Gispen WH. Modulation of brain polyphosphoinositide metabolism by ACTH-sensitive protein phosphorylation. *Nature* 1980;286: 623-5.
- 359. Van Dongen C, Zwiers H, Oesreicher AB, Gispen WH. ACTH, phosphoprotein B-50, and polyphosphoinositide metabolism in rat brain membranes. In: Horrocks LA, Kanfer JN, Porcellati G, eds. *Phospholipids in the Nervous System, Vol 2: Physiological Roles*. New York: Raven Press, 1985:49–59.
- 360. Holmes GL, Weber DA. Effects of ACTH on seizure susceptibility in the developing brain. Ann Neurol 1986;20:82-8.
- 361. Leonard BE. The effect of two synthetic ACTH analogues on the metabolism of biogenic amines in the rat. Arch Int Pharmacodyn 1974;20:242-53.
- 362. Versteeg DHG. Interaction of peptides related to ACTH, MSH and B-LPH with neurotransmitters in the brain. *Pharmacol Ther* 1980;11:535-57.
- 363. Kendall DA, McEwen BS, Enna SJ. The influence of ACTH and corticosterone on [³H]GABA receptor binding in rat brain. *Brain Res* 1982;235:365-74.
- 364. Chiriboga CA, Pranzatelli MR, De Vivo DC. The effect of ACTH treatment on striatal dopamine D-2 receptors in developing rat brain. Ann Neurol 1987;22:433.
- 365. Pranzatelli MR. In vivo and in vitro effects of adrenocorticotrophic hormone on serotonin receptors in neonatal rat brain. *Dev Pharmacol Ther* 1989;12:49–56.
- 366. Pranzatelli MR, Eng B. Chronic ACTH treatment: Influence on 5-HT₂ receptors and behavioral supersensitivity induced by 5,7-dihydroxytryptamine lesions. *Peptides* 1989;10:5–8.
- 367. Romagnano MA, Joseph SA. Immunocytochemical localization of ACTH₁₋₃₉ in the brainstem of the rat. *Brain Res* 1983;276:1–16.
- 368. Schwartzberg DG, Nakane PK. ACTH-related peptide containing neurons within the medulla oblongata of the rat. *Brain Res* 1983;276:351-6.
- Lewis DA, Sherman BM. Serotonergic stimulation of adrenocorticotropin secretion in man. J Clin Endocrinol Metab 1984;58:458–462.
- 370. Smith J, Fauquet M. Glucocorticoid stimulation of adrenergic differentiation in cultures of migrating and premigratory neural crest. J Neurosci 1984;4:2160-72.
- 371. Worlledge SM. Autoantibody formation associated with methyldopa (Aldomet) therapy. Br J Haematol 1969;16:5.
- 372. Berry-Kravis E, Dawson G. Modulation of an adenylate cyclase-linked serotonin (5-HT₁) receptor system in neuroblastoma X brain explant hybrid cell line (NCB-20) by opiates, prostaglandins, and α_2 -adrenergic agonists. J Neurochem 1983;40:361-72.
- 373. Berry-Kravis E, Dawson G. Characterization of an adenylate cyclase-linked serotonin (5-HT₁) receptor in a neuroblastoma X brain explant hybrid cell line (NCB-20). J Neurochem 1983;40: 977–85.
- 374. Chang K-J, Eckel RW, Blachard SG. Opioid peptides induce reduction of enkephalin receptors in cultured neuroblastoma cells. *Nature* 1982;296:446–8.
- 375. Gilbert JA, Richelson E. Influence of delta-opioid receptors on production of labeled methionine⁵-enkephalin in murine neuroblastoma cells. J Neurochem 1985;44:922-8.
- Glaser T, Hubner K, Hamprecht B. Neuroblastoma X glioma hybrid cells synthesize enkephalinlike opioid peptides. J Neurochem 1982;39:59–69.
- 377. McKinney M, Stenstrom S, Richelson E. Muscarinic responses and binding in a murine neuroblastoma clone (N1E-115). *Mol Pharmacol* 1985;27:223-35.
- 378. Sabol S, Nirenberg M. Regulation of adenylate cyclase of neuroblastoma X glioma hybrid cells by α -adrenergic receptors. I. Inhibition of adenylate cyclase mediated by α -receptors. J Biol Chem 1979;254:1913-20.
- 379. Traber J, Fischer K, Buchen C, Hamprecht B. Muscarinic responses to acetylcholine in neuroblastoma X glioma hybrid cells. *Nature* 1975;255:558–9.
- 380. Hoyer D, Neijt HC, Karpf A. Competitive interaction of agonists and antagonists with 5-HT₃ recognition sites in membranes of neuroblastoma cells labelled with [³H]ICS-205-930. J Recept Res 1989;9:65-9.
- 381. Reboulleau CP. Multiple types of neurotransmitter binding sites in the rat neuroblastoma B50 cell

line. I. Characterization and binding affinity changes during various differentiation paradigms. *Dev Brain Res* 1987;31:201-12.

- 382. MacDermot J, Higashida H, Wilson S, Matsuzawa H, Minna J, Nirenberg M. Adenylate cyclase and acetylcholine release regulated by separate serotonin receptors of somatic cell hybrids. Proc Natl Acad Sci USA 1979;76:1135–9.
- 383. Duasse J-P, Diop L. Monoclonal antibodies to rat brain α-adrenoreceptors. Eur J Pharmacol 1983;95:135-7.
- 384. Fraser CM, Venter JC. Monoclonal antibodies to β-adrenergic receptors: Use in purification and molecular characterization of β-receptors. Proc Natl Acad Sci USA 1980;77:7034–8.
- 385. Fazekas de St Groth, Schneidegger D. Production of monoclonal antibodies: Strategy and tactics. J Immunol Meth 1980;35:1–21.
- 386. Cuello AC. Monoclonal antibodies to neurotransmitter: Potential value in the understanding of normal and abnormal neurological function. In: McMichael AJ, Fabre JW, eds. *Monoclonal antibodies in clinical medicine*, New York: Academic Press; 1982:413-28.
- 387. Vincent A. Immunology of acetylcholine receptors in relation to myasthenia gravis. *Physiol Rev* 1980;60:7567–82.
- 388. Vincent FM. Paraneoplastic CNS and renal syndromes. Simultaneous occurrence in a patient with bronchogenic carcinoma. JAMA 1978;240:862–3.
- 389. Knight A, Adams DO. Autoantibodies with intrinsic biological activity. Horm Res 1980;13:69-80.
- 390. Solimena M, Folli F, Denis-Donini S, et al. Autoantibodies to glutamic acid decarboxylase in a patient with stiff man syndrome, epilepsy and type I diabetes mellitus. N Engl J Med 1988;318: 1012-20.
- Schuller-Petrovic S, Gebhart W, Lassman H, Rumpold H, Kraft D. A shared antigenic determinant between natural killer cells and nervous tissue. *Nature* 1983;306:179–81.
- 392. Pranzatelli MR, Balletti J. Characterization of 5-hydroxytryptamine 1A-like binding sites in human ganglioneuroblastoma. *Neurosci Lettt* 1991;132:117-20.
- 393. Pranzatelli MR, Balletti J, Levy M. Neurotransmitter receptors on human neural crest tumors: A new hypothesis for the opsoclonus-myoclonus syndrome [Abstract]. Ann Neurol 1989;26:474.
- Pranzatelli MR, Balletti J. Identification of 5-HT_{1A} recognition sites in human ganglioneuroblastomas. Eur J Pharmacol 1989;170:127-8.
- 395. Tricklebank MD. The behavioral response to 5-HT receptor agonists and subtypes of the central 5-HT receptor. *Trends Pharmacol Sci* 1985;14:403–7.
- 396. Pranzatelli MR, Balletti J. Serotonin receptors in human neuroblastoma: A possible biologic tumor marker. *Exp Neurol* 1992;115:423-7.
- 397. Van Woert MH, Hwang EC. Role of brain serotonin in myoclonus. In: Morselli PL, Lloyd KG, Loscher W, Meldrum B, Reynolds EH, eds. *Neurotransmitters, seizures, and epilepsy*, New York: Raven Press, 1981:239–49.
- 398. Chadwick D, Hallett M, Harris R, Jenner P, Reynolds EH, Marsden CD. Clinical, biochemical and physiological features distinguishing myoclonus responsive to 5-hydroxytryptophan, tryptophan with monoamine oxidase inhibitor, and clonazepam. *Brain* 1977;100:455–87.
- 399. Pranzatelli MR. Effect of chronic treatment with 5-hydroxytryptophan on cortical serotonin receptors in the rat. *Clin Neuropharmacol* 1988;11:257-62.
- 400. Todd RD, Ciaranello RD. Demonstration of inter- and intraspecies differences in serotonin binding sites by antibodies from an autistic child. *Proc Natl Acad Sci USA* 1985;82:612–6.
- 401. Trifilletti RR, Pranzatelli MR. The peripheral-type benzodiazepine receptor in a series of neuroectodermal tumors of childhood [Abstract]. *Neurology* 1990;(40) Suppl:361.
- 402. Julius D, Livelli JJ, Jessel TM, Axel R. Ectopic expression of the serotonin_{1C} receptors and the triggering of malignant transformation. *Science* 1989;244:1057–62.
- Chelmicka-Szorc E, Arnason BGW. Effect of 6-hydroxydopamine on tumor growth. Cancer Res 1976;36:2382–4.
- 404. Wigglesworth VB. A case of DDT poisoning in man. Br Med J 1945;14:517.
- 405. Haymaker E, Ginzler AM. The toxic effects of prolonged ingestion of DDT on dogs with special reference to lesions in the brain. Am J Med Sci 1986;212:423-31.
- 406. Van Woert MH, Hwang EC. Animal models of myoclonus. Adv Neurol 1979;26:173-9.
- 407. Chung E, Van Woert MH. DDT myoclonus: Sites and mechanism of action. *Exp Neurol* 1984; 85:273-82.
- 408. Gandolfi O, Cheney DL, Hong J-S, Costa E. On the neurotoxicity of chlordecone: A role for γ-aminobutyric acid and serotonin. *Brain Res* 1984;303:117-23.
- 409. Roba J, Cavalier R, Cordi A, et al. Milacemide. In: Meldrum BS, Porter RJ, eds: New anticonvulsant drugs. Current problems in epilepsy IV, London: John Libbey, 1986:179-90.

- 410. Van Dorsser W, Barris D, Cordi A, Roba J. Anticonvulsant activity of milacemide. Arch Int Pharmacodyn Ther 1983;266:239-249.
- 411. Hunter C, Chung E, Van Woert MH. Antimyoclonic action of milacemide in p,p'-DDT-induced myoclonus in mice. Soc Neurosci Abstr 1986;12:1243.
- 412. Lisi L de. Su di un fenomeno motorio costante del sonno normale: Le mioclonie ipniche fisiologische. I. Descrizione. [On a constant motor phenomenon of normal sleep: Physiological hypnotic myoclonia] Riv Patol Nerv Ment 1932;39:481-96.
- 413. Pompeiano O. The generation of rhythmic discharges during bursts of REM. In: Chalazonitis N, Boiusson M, eds. Abnormal neuronal discharges. New York: Raven Press, 1978.
- 414. Jouvet M. Neurophysiology of the states of sleep. Physiol Rev 1967;47:117-77.
- 415. Jacobs L, Feldman M, Bender MB. Eye movements during sleep. I. The pattern in the normal human. Arch Neurol 1971;25:151-9.
- 416. Mahowald MU, Schenck CH. REM sleep behavior disorder. In: Kryger MH, Roth T, Dement WC, eds. Principles and practice of sleep medicine. Philadelphia: Saunders, 1989:389-401.
- Denny-Brown D. Quelques aspects physiologiques des myoclonies. [A few physiological aspects of myoclonia] In: Bonduelle M, Gastaut H, eds. Les myoclonies. Paris: Masson, 1968:121-9.
- 418. Mano T, Shaiozawa Z, Sobue I. Extrapyramidal involuntary movements during sleep. In: Broughton RJ, ed. Henry Gastaut and the Marseilles school's contribution to the neurosciences (EEG suppl 35). Amsterdam: Elsevier Biochemical Press, 1982;431-42.
- 419. Silvestri R, De Domenico P, Di Rosa AE, Bramanti P, Serra S, Di Perri R. The effect of nocturnal physiological sleep on various movement disorders. *Mov Disord* 1990;5:8–14.
- 420. Chase MH, Morales FR. The atonia and myoclonia of active (REM) sleep. Annu Rev Psychol 1990;41:557-84.
- Steriade M, Hobson JA. Neuronal activity during the sleep-waking cycle. *Prog Neurobiol* 1976; 6:155–376.
- 422. Lai YY, Siegel JM. Medullary regions mediating atonia. J Neurosci 1988;8:4790-6.
- 423. Mori S, Ohta Y, Takakusaki K, Matsuyama K, Sugaya K. Pontomedullary and spinal mechanisms of postural suppression in a decerebrate, reflex standing cat. In: Novel approaches to the study of motor systems, Satellite Symp., 30th Int. Cong. Physiol. Sci., Banff, Alberta, Canada, July 10-13, 1986.
- 424. Sakai K, Sastre JP, Kanamori N, Jouvet M. State-specific neurons in the ponto-medullary reticular formation with special reference to the postural atonia during paradoxical sleep in the cat. In: Pompeiano O, Ajmone Marsan C, eds. *Brain mechanisms and perceptual awareness*, New York: Raven Press, 1981.
- 425. Lai YY, Siegel JS. Medullary control of locomotion and myoclonic jerks [Abstract]. Soc Neurosci 1991;187:15.
- Fuchs AF, Kaneko CRS, Seudder CA. Brainstem control of saccadic eye movements. Annu Rev Neurosci 1985;8:307–37.
- 427. Zee DS, Robinson DA. A hypothetical explanation of saccadic oscillations. Ann Neurol 1979;5: 405-21.
- 428. Bergenius J. Saccade abnormalities in patients with ocular flutter. Acta Otolaryngol 1986;102: 228-33.
- 429. Ellenberger C Jr, Keitner JL, Stroud MH. Ocular dyskinesia in cerebellar disease. Brain 1972; 95:685-92.
- 430. Dichgans J, Jung R. Oculomotor abnormalities due to cerebellar lesions. In: Lennerstrand G, Bach-y-Rita P, eds. *Basic mechanisms of ocular motility and their clinical implications*, New York: Pergamon Press, 1975:285–7.
- 431. Goldberg RT, Jampel RS. Flutter-like oscillations of the eyes in cerebellar disease. Am J Ophthalmol 1963;55:1229-33.
- 432. Wertenbaker C, Behrens MM, Hunter SB, Plank CR. Opsoclonus—a cerebellar disorder? Neuro-Ophthalmology 1981;2:73-84.
- 433. Gilbert GJ, McEntee WJ III, Glaser GH. Familial myoclonus and ataxia. *Neurology* 1963;13: 365-72.
- Jayakar PB, Seshia SS. Involuntary movements with cerebellar tumour. Can J Neurol Sci 1987; 14:306–8.
- 435. Mettler FA. Supratentorial mechanisms influencing the oculomotor apparatus. In: Bender MB, ed. *The oculomotor system*, New York: Paul B Hoeber, 1964:1-17.
- 436. Zee DS, Optician LM. Mechanisms of ocular oscillations. In; Findley LJ, Capildeo R, eds. Movement Disorders: Tremors, New York: Oxford University Press, 1984:409-19.
- 437. Ridley A, Kennard C, Scholtz CL, Buttner-Ennever JA, Summers B, Turnbull A. Omnipause

neurons in two cases of opsoclonus associated with oat cell carcinoma of the lung. Brain 1987; 110:1699-709.

- 438. Hunter S, Kooistra C. Neuropathologic findings in idiopathic opsoclonus and myoclonus. Their similarity to those in paraneoplastic cerebellar cortical degeneration. J Clin Nerv Ophthalmol 1986;6:236-41.
- 439. Tychsen L [cited by Digre 102]. Methyltryosine produces square-wave jerks in healthy humans testing a model of pause cell control [Dissertation]. Ames, Iowa, University of Iowa, 1981.
- 440. Baloh RW, Dietz J, Spooner JW. Myoclonus and ocular oscillations induced by L-tryptophan. Ann Neurol 1982;11:95-7.
- 441. Hikosaka O, Wurtz RH. Modification of saccadic eye movements by GABA-related substances.
 II. Effects of muscimol in monkey substantia nigra pars reticula. J Neurophysiol 1985;53:292–308.
- 442. Giordana MT, Soffietti R, Schiffer D. Paraneoplastic opsoclonus: A neuropathologic study of two cases. *Clin Neuropathol* 1989;8:295-300.
- 443. Verhaart WJC. Grey matter degeneration of the central nervous system in carcinosis. Acta Neuropathol 1961;1:107-12.
- 444. Vick N, Schulman S, Dan P. Carcinomatous cerebellar degeneration, encephalomyelitis, and sensory neuropathy (radiculitis). *Neurology* 1969;19:425–41.
- 445. Dorfman LJ, Forna LS. Paraneoplastic encephalomyelitis. Acta Neurol Scand 1972;48:556-73.
- 446. Reddy RV, Vakili JT. Midbrain encephalitis as a remote effect of a malignant neoplasm. Arch Neurol 1981;38:781-2.
- 447. Howell DA, Lees AJ, Toghill PJ. Spinal internuncial neurones in progressive encephalomyelitis with rigidity. J Neurol Neurosurg Psychiatry 1979;42:773-85.
- 448. Boddie HG. Ocular bobbing and opsoclonus. J Neurol Neurosurg Psychiatry 1972;35:739-42.
- 449. Lalonde R, Botez MI. The cerebellum and learning processes in animals. Brain Res Rev 1990; 15:325-32.
- 450. Marr D. A theory of cerebellar cortex. J Physiol 1969;202:437-70.
- 451. Llinas R, Walton K, Hillman DE, Sotelo C. Inferior olive: Its role in motor learning. Science 1975;190:1230-1.
- 452. Botez MI, Botez T, Elie R, Attig E. Role of the cerebellum in complex human behavior. Ital J Neurol Sci 1989;10:291-300.
- 453. Hamilton NG, Frick RB, Takahasi T, Hopping MW. Psychiatric symptoms and cerebellar pathology. Am J Psychiat 1983;140:1322-6.
- 454. Bauman M, Kemper TL. Histoanatomic observations of the brain in early infantile autism. Neurology 1985;35:866-74.